

DISEASE NOTE

FIRST REPORT OF GALL DISEASE IN MANGO TREES CAUSED BY *FUSARIUM DECEMCELLULARE* IN DOMINICAN REPUBLIC

E. García-López¹, J.A. Mora-Aguilera¹, E. Hernández-Castro², C.J. Jiménez-Vásquez³, C.M. Batista-Martínez⁴ and C. Serra⁴

¹Fitopatología, Instituto de Fitosanidad, Campus Montecillo, Colegio de Postgraduados, km 36.5 Carretera México-Texcoco, 56230, Texcoco, Estado de México, México

²Sistemas de Producción Agropecuaria, Universidad Autónoma de Guerrero, km. 2.5 carretera Iguala-Tuxpan, 40101, Iguala, Guerrero, México

³Departamento de Desarrollo de Frutales, Ministerio de Agricultura, Jardines del norte km 61/2 autopista Duarte, 745, Santo Domingo oeste, República Dominicana

⁴Estación Experimental de Frutales Bani, Instituto Dominicano de Investigaciones Agropecuarias y Forestales, Villa Sombrero, 94000, provincia Bani, República Dominicana

Malformed vegetative shoots were collected in mango orchards cv. 'Puntica' in Peravia, province of Dominican Republic during July-September 2013, the incidence ranged from 10 to 50%. *Fusarium* were consistently isolated on potato-dextrose-agar media. After 14 days of incubation at 28°C colonies developed intense rose pigmentation, macroconidia were 7-8 septate, 45.9-76.9×5.4-6.7 µm, with rounded apical cell and footlike basal cell; microconidia were ovoid, 0-1 septate, 5.3-12.4×2.0-4.6 µm. Based on morphology, the fungus was identified as *Fusarium decemcellulare* (Leslie and Summerell, 2006). DNA was extracted from a representative monospore culture (EEFB-1). Elongation factor 1-alpha and beta-tubulin gene regions were amplified using EF1/EF2 and BT1/BT2 primers. PCR products were purified and sequenced. The obtained sequences of 683 and 193 bp were deposited in the GenBank (Accession Nos. KX345392 and KX345393). BLASTn analysis showed 99 and 86% homology with *F. decemcellulare* sequences HM054059 and HM054098. Pathogenicity tests were conducted on ten healthy 6-month-old mango plants cv. 'Puntica', by infiltration of 30 µl of a conidial suspension (2×10⁶ spores/ml) into apical bud. Ten control plants were infiltrated with sterile water. Inoculated shoots with *F. decemcellulare* showed excessive proliferation of axillary buds that remained protuberant or stunted and coriaceous, 8-9 months after inoculation. The fungus was reisolated only from diseased shoots, fulfilling Koch's postulates. Although *F. decemcellulare* has been previously reported causing mango's galls in Mexico, USA, Brazil and Puerto Rico (Angulo and Villapudua, 1982; Farr and Rossman, 2016), this is the first report of this pathogen on mango in Dominican Republic.

Angulo S.M., Villapudua J.R. 1982. Buba of mango (*Mangifera indica* L.) in the state of Sinaloa, Mexico. *Phytopathology*: 72: 171.

Farr D.F., Rossman A.Y. Fungal Databases, Systematic Mycology and Microbiology Laboratory, ARS, USDA. Retrieved June 23, 2016, from <http://nt.ars-grin.gov/fungalDATABASES/>

Leslie J.F., Summerell B.A., 2006. The *Fusarium* Laboratory Manual. Blackwell Publishing. Ames, Iowa, USA.

Corresponding author: J.A. Mora-Aguilera
E-mail: aguilera@colpos.mx

Received July 18, 2016
Accepted November 10, 2016

DISEASE NOTE

FIRST REPORT OF POWDERY MILDEW CAUSED BY *OIDIUM HELIOTROPII-INDICION HELIOTROPIUM INDICUM* IN INDIA

A.K. Nayak and B.K. Babu

Microbial Genomics and Diagnostics Lab., Microbiology and Plant Pathology Division, Regional Plant Resource Centre, Bhubaneswar-751015, Odisha, India

Indian heliotrope (*Heliotropium indicum* L., family Boraginaceae) is an annual, herbaceous plant and a common weed in waste lands areas. During field surveys conducted between September to January 2015 a severe powdery mildew outbreak was observed in different areas of the Odisha State (India). Symptoms initially appeared as small circular to irregular white spots on both leaf sides, which expanded resulting in plant defoliation and drying. Conidia scraped from symptomatic leaves were ellipsoid to ovoid, had distinct fibrosin bodies and were 23-35×13-20 µm in size (l/w ratio 1.4-2.1). Conidiophores were erect, straight, slightly curved, cylindrical, foot cell 43 to 63×12 to 18 µm, followed by 1-3 shorter hyaline cells. Germ tubes stemmed from the lateral sides of the conidia. Based on the above, the fungus was identified as *Oidium heliotropii-indici* (Braun and Cook, 2002) in the *Podosphaera* section, *Sphaerotheca* subsection (Cook and Braun, 2009). *O. heliotropii-indici* and the anamorphic state of *Podosphaera fuliginea* cannot be easily distinguished on the basis of the appressorium morphology (Braun, 1987) and the main difference is the nipple shape in *O. heliotropii-indici* in contrast to the indistinct shape in *P. fuliginea*. Pathogenicity was confirmed by gently pressing an infected leaf on to the young leaves of five healthy potted *H. indicum* plants. Five non-inoculated plants were used as controls. Plants were maintained in a greenhouse at 28 to 30°C. Seven to ten days post inoculation, typical white patches appeared on the upper side of the inoculated leaves; they were morphologically similar to those observed on naturally infected plants whereas control plants remained symptomless. To the best of our knowledge, this is the first confirmed report of *O. heliotropii-indici* causing powdery mildew on *H. indicum* in India.

Braun U., 1987. A monograph of the Erysiphales (powdery mildews). Beih. *Nova Hedwigia* 89: 1-700.

Braun U., Cook R.T.A., Inman A.J., Shin H.D., 2002. The taxonomy of the powdery mildew fungi. In: Bélanger R.R., Bushnell W.R. (eds). The Powdery Mildews: a Comprehensive Treatise, pp. 13-55. APS Press, St. Paul, MN, USA.

Cook R.T.A., Braun U., 2009. Conidia germination patterns in powdery mildews. *Mycological Research* 113: 616-636.

Corresponding author: B.K. Babu
E-mail: kishore_bandam@yahoo.co.in

Received August 29, 2016
Accepted February 8, 2017