

DISEASE NOTE

FIRST REPORT OF JUJUBE DIEBACK
CAUSED BY *FUSARIUM SOLANI*M.R. Mirzaee¹, M. Jahani², H. Mahmoudi¹ and K. Ghos³¹ Agricultural and Natural Resources Research Center
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In spring 2010, a severe branch dieback was observed in jujube (*Ziziphus jujuba*) orchards of the Sarbisbah region of southern Khorasan (Iran). Symptoms included twig dieback, blackish discoloration of wood and foliage, and wilting followed by leaf shedding. Samples of symptomatic tissues, including discoloured wood and cambium from twigs and branches, were excised, surface-sterilised, plated onto Petri dishes with potato dextrose agar (PDA), and incubated in the dark at 25°C for 10 days. On PDA, the colonies presented a cream-coloured mycelium with yellow pigmentation and a creamy reverse. On carnation leaf agar, macroconidia were abundant, thick walled, 4 to 6 celled, hyaline, fusoid, and measured 5-7×20-37.5 µm. Microconidia, which were formed in false heads developing on long monophialides, were oval, reniform and fusiform, had 0 to 2 septa and measured 2.5-5×7-15 µm. Chlamydospores were single, double or several in chain, and measured 5-7.5×5-9 µm. Based on these morphobiometric and cultural characteristics, the fungus was identified as *Fusarium solani* (Demicri and Maden, 2006; Leslie and Summerell, 2006). For pathogenicity tests, normal and wounded shoots of three 6-year-old jujube trees were inoculated with 5 mm diameter mycelium disks from two 10-day-old single-spore isolates. Inoculation sites were covered with moistened cotton wool and parafilm. Control inoculations were made with sterile PDA. Disease symptoms, including tissue discoloration, were first observed 14 days after inoculation of the shoots, from which the fungus was reisolated. Controls did not develop symptoms. *F. solani* is frequently found associated with damage or stress events caused by other biotic or abiotic agents (Booth, 1971). This represents the first evidence of *F. solani* pathogenicity to *Z. jujuba*.

Booth C., 1971. The Genus *Fusarium*. Commonwealth Mycological Institute, Kew, UK.Demicri F., Maden S., 2006. A severe dieback of box elder (*Acer negundo*) caused by *Fusarium solani* (Mart.) Sacc. in Turkey. *Australasian Plant Disease Notes*, 1:13-15.Leslie J.F., Summerell B.A., 2006. The *Fusarium* Laboratory Manual. Blackwell Publishing, Oxford, UK.

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FIRST REPORT OF *PLUM BARK
NECROSIS STEM PITTING-ASSOCIATED
VIRUS* IN STONE FRUIT TREES
IN TUNISIAW. Salleh^{1,2}, N. Mahfoudhi² and K. Djelouah¹¹ Istituto Agronomico Mediterraneo, Via Ceglie 9,
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During a survey carried out in two stone fruit mother blocks located at Kairouan (central Tunisia) and Jendouba (northern Tunisia), a total of 212 leaf samples were collected from peach (70), almond (37), apricot (53), plum (36) and cherry (16) trees, respectively. These samples were tested by DAS-ELISA for the presence of *Plum bark necrosis stem pitting-associated virus* (PBNSPaV) using specific antibodies (Agritest, Italy) according to the manufacturer's instructions. Serological analysis showed that nearly 8% of the samples were infected by PBNSPaV, particularly those from peach (15.7%), plum (11.1%) and almond (5.4%) trees. Virus presence was confirmed by RT-PCR using the specific primers ASP1 5' CGGTAGGGCTGTGACTACCG-3' and ASP2 5'-GTAGTCCGCTGGTACGCTCAAG-3' (Abou Ghanem-Sabanadzovic *et al.*, 2001) and total RNA extracted from leaf veins according to Foissac *et al.* (2001). This assay confirmed the results obtained by serology and identified three more infected samples, one from peach and two from apricot, which were negative in DAS-ELISA. None of the infected trees showed particular field symptom. To the best of our knowledge, this is the first report of PBNSPaV from Tunisia.

Abou Ghanem-Sabanadzovic N., Mahboubi M., Di Terlizzi B., Sabanadzovic S., Savino V., Uyemoto J.K., Martelli G.P., 2001. Molecular detection of a closterovirus associated with apricot stem pitting in southern Italy. *Journal of Plant Pathology* 83: 125-132.Foissac X., Svanella-Dumas L., Gentit P., Dulucq M.J., Candresse T., 2001. Polyvalent detection of fruit tree Tricho-, Capillo- and Foveaviruses by nested RT-PCR using degenerated and inosine containing primers (DOP RT-PCR). *Acta Horticulturae* 550, 37-43.

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