



Review Article

Organisms associated with the family Convolvulaceae and their potential for biological control of *Convolvulus arvensis*

Peter Tóth* and Ľudovít Cagán

Department of Plant Protection, Slovak Agricultural University, A. Hlinku 2, 949 76 Nitra, Slovak Republic

*Author for correspondence: petery@nexta.sk

Abstract

This review catalogues the organisms that have been recorded in association with the family Convolvulaceae worldwide. It selects promising organisms and provides basic information on these potential biological control agents. In total, 328 organisms associated with the Convolvulaceae have been reported in the 270 references cited. Wasps, bees and flies are excluded where they are only casual and transient visitors to the plants. Although a relatively large number of species has been recorded from the Convolvulaceae, only a small number (47 species, 14.3%) seem to have the potential for biological control of *Convolvulus arvensis*. Of these, *Melanagromyza albocilia* (Dipt., Agromyzidae), *Longitarsus pellucidus* and *Hypocassida subferruginea* (Col., Chrysomelidae), *Spermophagus sericeus* (Col., Bruchidae), *Emmelia trabealis* (Lep., Noctuidae) and *Emmelina monodactyla* (Lep., Pterophoridae) appear to be the most promising.

Keywords: Field bindweed: *Convolvulus arvensis*: Convolvulaceae: phytophagous insects: biological control: agent selection

1. Introduction

Field bindweed (*Convolvulus arvensis* L.) and hedge bindweed (*Calystegia sepium* (L.) R. Br.), both from the family Convolvulaceae, are very important weeds in world agriculture. They are deep-rooted perennials and thus they can escape chemical or mechanical weed control [1, 2]. In addition, the abundant reserves stored in the roots enable the plants to survive repeated defoliation [3]. Moreover, fragmentation and dispersal of underground parts leads to vegetative propagation [4]. In this context, biological control appears to be a promising alternative strategy against bindweeds.

Bindweed relatives in the Convolvulaceae include *Ipomoea batatas* (L.) Lam. (sweet potato) and ornamental species in the genus *Ipomoea*. Sweet potato, a widely grown plant, is a staple food for much of the population in many parts of the world [5]. Because sweet potato and some ornamental *Ipomoea* species are often subjected to attack by insects associated

with bindweeds, conflicting interests is the major problem to solve in the search for biological control agents for bindweeds [6]. Species feeding on sweet potato have value for biological control of bindweeds only in areas with no significant production of sweet potato [7]. However, any organisms which curtail weed growth or reproduction may be used as biological control agents [8]. Thus, the search for natural enemies should encompass all organisms associated with the target plants [9], including pathogens [10].

The first step in any biological weed control programme is a comprehensive literature survey [11, 12]. Mohyuddin [13] listed 125 species of insects and mites known to attack *Convolvulus* spp. and *Calystegia* spp., found by him in Canada and by others throughout the world. Thirty-five species of insect and two species of mite were found in association with *Convolvulus* spp. in Pakistan [14]. The insect fauna noted on *C. arvensis* in Egypt consisted of 13 species [15]. In Mediterranean Europe, 139 species of phytophagous arthropods were collected from *C.*

arvensis and its close relatives [5]. Weaver & Riley [1] summarized organisms collected from *C. arvensis* in Canada and the USA in a list of 28 insect species. Field surveys of phytophagous insects associated with *Convolvulus* spp. were also carried out in Slovakia to evaluate their potential use in biological control programmes. During the study 108 organisms were collected from *C. arvensis* [16]. Other authors highlighted promising biological control candidates: Mohyuddin [17] in Canada, Baloch [18, 19] in Pakistan, Tyurebaev [20] in Central Asia, Gorenshstein [21] in Russia, and Wang & Kok [7] and Julien [22] worldwide.

This review catalogues all organisms that have been recorded from the Convolvulaceae worldwide. It then identifies species with potential for use in biological control of bindweeds, particularly *C. arvensis*. It summarizes available biological information for each of these species, and discusses the prospects for and limitations to their use in biological control programmes.

2. Organisms Associated with the Family Convolvulaceae

Organisms which have been recorded on Convolvulaceae worldwide are listed in Table 1. Wasps, bees and flies where they are only casual and transient visitors to the plants have been excluded. In total 328 organisms associated with the Convolvulaceae have been recorded. These represent ten orders and 58 families. The majority of species belong to the insect orders Coleoptera (43.7% species), Lepidoptera (33.9%) and Hemiptera (13.6%) (Table 2). Numbers of species and families represented in each order are also given in Table 2.

The Palaearctic fauna (46.8% species) was noticeably richer compared to other regions of the world. The greatest diversity of insect fauna associated with *C. arvensis* was found in Eurasia and North Africa, reflecting the plant's Eurasian origin. On the other hand, organisms feeding on sweet potato were reported from tropical and subtropical regions. The exact origin of the sweet potato is not known, but a New World origin is generally accepted [251]. This is probably why the majority (more than 56%) of species feeding on sweet potato were recorded in the Americas.

The most commonly listed feeding habit was external or internal feeding on leaves (about 54.0% of the 328 recorded species). This was followed by feeding on roots (8.0%), seeds (4.9%), flowers (3.3%) and stems (3.3%). The feeding habit of 87 species (26.5%) was not specified in the literature (Table 3).

The organisms included in this review may be divided into two main groups: polyphagous species (45.7%), and species whose feeding was restricted to the Convolvulaceae (54.3%). There seemed to be just 22 strictly monophagous species (6.5%), feeding on

only *C. arvensis* or *Calystegia sepium*. A group of about 47 species (14.3%) was specific to *Convolvulus* spp. A group of oligophagous insects (feeding on a relatively large number of plants in different genera within the Convolvulaceae) contained 59 species (17.6%). Sweet potato was reported as a host for 56 species (16.6%).

3. Potential Biological Control Agents

Although a relatively large number of species has been recorded from the Convolvulaceae, only a small number of them (47 species, 14.3%) seem to have potential as biological control agents. Of these, only *Tyta luctuosa* (Denis & Schiffermüller) (Lep., Noctuidae) and *Aceria malherbae* Nuzzaci (Acari, Eriophyidae) have so far been used in the classical biological control of bindweeds [33].

This estimate of the number of potential biological control agents is probably not entirely accurate. It can be difficult to determine whether or not a species is suitable for biological control. For example, *Longitarsus pellucidus* (Foudras) (Col., Chrysomelidae) has been recorded as a polyphagous species [96]. However, feeding tests have shown that it fed only on species in the Convolvulaceae [16].

On the basis of field surveys and laboratory testing, 12 species were selected as promising biological control agents in Slovakia [16]. Rosenthal & Buckingham [5] selected nine promising species for the same purpose in Mediterranean Europe, and Baloch [19] identified four species in Pakistan. Wang & Kok [7] stated that there were seven promising candidates worldwide. However, according to the feeding tests conducted by Mohyuddin [13], none of the leaf-feeding insects studied was suitable for biological control in sweet potato growing areas.

Within families, the species identified as promising tended to be those attacking the roots or stems. For example, the species of Cerambycidae (Coleoptera), Sesiidae (Lepidoptera) and Cecidomyiidae and Agromyzidae (Diptera) noted to have the most potential as biological control agents all fed on roots or shoots. Given the tremendous amount of root material produced by *C. arvensis*, it is surprising that more organisms do not utilize roots as a food source. Species of Bruchidae (Coleoptera) were suggested as prospective agents for their ability to destroy seeds and seedlings in five of eight studies dealing with potential biological control agents [1, 5, 7, 16, 20]. Flea beetles and tortoise beetles (Chrysomelidae), whose adults are leaf feeders, were suggested as promising species for biological control purposes in 16.5% of 79 studies, because of their restricted host range, larval damage concentrated in the roots or leaves and high adaptability to different ecological situations. Lepidopteran and coleopteran leaf-feeding species were suggested as potential biological control agents usually only if they were monophagous. The promising species are identified in Table 1.

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
NEMATODA					
TYLENCHATA					
Tylenchidae					
<i>Ditylenchus dipsaci</i> (Kühn) Filipjev	Europe	EXT, R	POLY		23
Heteroderidae					
<i>Meloidogyne hapla</i> Chitwood	Germany	EXT, R	POLY		24
<i>Meloidogyne incognita</i> ssp. <i>acrita</i> (Kofoid & White) Chitwood	South Africa, Egypt, Nigeria	EXT, R	CONVOL		25, 26, 27
<i>Meloidogyne javanica</i> (Treub) Chitwood	Egypt, South Africa	EXT, R	POLY, IB		25, 27
<i>Meloidogyne</i> sp.	Europe	EXT, R	POLY		23
Hoplolamidae					
<i>Rotylenchulus reniformis</i> Linford & Oliveira	India	EXT, R	POLY		28
ARACHNOIDEA					
ACARI					
Eriophyidae					
<i>Aceria malherbae</i> Nuzzaci	Europe, North Africa	EXT, L, St	CC	*	23, 29 – 33
<i>Eriophyes altus</i> Nalepa [<i>Aceria altus</i>]	Indonesia	EXT, L	ID		30
<i>Eriophyes gastrotrichus</i> Nalepa [<i>Aceria gastrotrichus</i>]	Philippines	EXT, L	IB, IS		30, 34
<i>Eriophyes lepidemosis</i> Nalepa [<i>Aceria lepidemosis</i>]	Indonesia	EXT, L	LB		30
<i>Eriophyes merremiae</i> Nalepa [<i>Aceria merremiae</i>]	Indonesia	EXT, L	MG		30
<i>Eriophyes</i> sp.	Italy, Spain	EXT, L	C		5
<i>Phyllocoptes convolvuli</i> (Nalepa) [<i>Aceria convolvuli</i>]	Europe	EXT, L	CA		23, 30
<i>Tegonotus convolvuli</i> (Channabasavanna)	India	EXT, L	I		30, 35
Tetranychidae					
<i>Tetranychus cinnabarinus</i> Boisduval	Italy	EXT, L	POLY		36
<i>Tetranychus urticae</i> Koch	Canada, Italy, South Africa, Poland	EXT, L	POLY		13, 27, 36, 37
INSECTA					
COLLEMBOLA					
Sminthuridae					
<i>Sminthurus viridis</i> (L.)	Italy	EXT, L	POLY		38
ORTHOPTERA					
Acrididae					
<i>Syrbula admirabilis</i> (Uhler)	–	–	POLY		13
Gryllidae					
<i>Gryllus burdigalensis</i> Latreille [<i>Modicogryllus burdigalensis</i>]	–	–	POLY		13
<i>Melanogryllus desertus</i> Pallas	–	–	POLY		13
THYSANOPTERA					
Thripidae					
<i>Scirtothrips aurantii</i> Faure	–	–	POLY		13

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon^b	Location	Plant association^c	Host specificity^d	Biocontrol candidate^e	Ref.
<i>Scolothrips</i> sp.	Italy	EXT, L	C		5
<i>Thrips tabaci</i> Lindeman	Egypt, Italy	EXT, L,F	POLY		15, 39
HEMIPTERA: HETEROPTERA					
Pentatomidae					
<i>Aelia acuminata</i> (L.)	Spain	EXT, SC	POLY		40
<i>Carpocoris mediterraneus</i> Tamanini	Spain	EXT, SC	POLY		40
<i>Cosmopepla bimaculata</i> Thomas	Canada	EXT, L	CONVOL		13
<i>Dryoderes umbraculatus</i> (F.)	Spain	EXT, SC	POLY		40
<i>Eurydema oleraceum</i> (L.)	Spain	EXT, SC	POLY		40
<i>Euschistus variolarius</i> (Palisot de Beauvois)	Canada	EXT, L	CONVOL		13
<i>Holcostethus strictus</i> (F.)	Spain	EXT, SC	POLY		40
<i>Nezara viridula</i> (L.)	Spain, Italy, Egypt	EXT, L, SC	POLY, CP		15, 39
Tingidae					
<i>Corythucha</i> sp.	Canada	EXT, L	CONVOL		13
Miridae					
<i>Adelphocoris lineolatus</i> (Goeze)	Canada	EXT, L	CONVOL		13
<i>Halticus tibialis</i> Reuter	Papua New Guinea	EXT, L	IB		41
<i>Lygus lineolaris</i> Palisot de Beauvois	USA	EXT, L	CONVOL		42
<i>Poecilocapsus lineatus</i> (F.)	Canada	EXT, L	CONVOL		13
HEMIPTERA: AUCHENORRHYNCHA					
Cixiidae					
<i>Hyalesthes obsoletus</i> Signoret	France, Germany, Slovakia	EXT, R, L	POLY		43 – 46
Delphacidae					
<i>Asiraca clavicornis</i> (F.)	Austria, Yugoslavia	EXT, L, St	POLY		47
Cercopidae					
<i>Cercopis sanguinolenta</i> (Scopoli)	Yugoslavia	EXT, L, F	POLY, CP		48
<i>Philaenus spumarius</i> L.	Europe, Canada	EXT, St	POLY, CP		13, 23, 49
Membracidae					
<i>Ceresa bubalus</i> F.	–	–	POLY		13
Cicadellidae					
<i>Cicadella viridis</i> (L.)	Italy	EXT, St	POLY, CP		48
<i>Empoasca</i> spp.	Peru	EXT, L	IB		50
<i>Petalcephala</i> sp.	–	–	POLY		13
<i>Platymetopius</i> sp.	Spain	EXT, St	C		5
<i>Selenocephalus pallens</i> Lindberg	Italy, Yugoslavia	EXT, St	POLY		5, 51, 52
HEMIPTERA: STERNORRHYNCHA					
Psyllidae					
<i>Trioza</i> sp.	Italy	EXT, L	C		5
Aleyrodidae					
<i>Aleyrodes spiraeoides</i> Quaintance	–	–	POLY		13
<i>Bemisia tabaci</i> (Gennadius)	Egypt, Uganda	EXT, L	POLY, CP		13, 15, 53, 54
Aphididae					
<i>Aphis convolvuli</i> Kaltentbach [<i>Myzus persicae</i> Sultzner]	Europe	EXT, L	CA		52

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Aphis fabae</i> ssp. <i>euonymi</i> F.	–	–	POLY		13
<i>Aphis fabae</i> Scopoli	Europe	EXT, L	POLY		23
<i>Aphis gossypii</i> Glover	USA, Canada	EXT, L	POLY		1, 34
<i>Aphis nerii</i> Boyer de Fonscolombe	USA, Canada	EXT, L	POLY		1
<i>Aphis spiraeicola</i> Patch	–	–	POLY		13
<i>Aulacorthum solani</i> (Kalenbach)	Yugoslavia	EXT, L	POLY, CP		47
<i>Aulacorthum speyeri</i> Börner	Italy	EXT, L	Convall., C		5, 55
<i>Macrosiphum euphorbiae</i> Thomas	Canada, Peru, Europe	EXT, L	POLY, CP		13, 39, 50, 56
<i>Macrosiphum gei</i> (Koch)	–	–	POLY		13
<i>Macrosiphum ludoviciana</i> e Oestlund [<i>Macrosiphoniella ludoviciana</i>]	–	–	POLY		13
<i>Medoralis althae</i> Kittel [<i>Aphis lavaterae</i>]	Europe	–	POLY		56
<i>Myzus persicae</i> Sultzer	Europe, Egypt, Canada	EXT, L	POLY, CP		13, 23, 34, 39, 56
<i>Rhopalosiphoninus latysiphon</i> Davidson	–	–	POLY		13
Ortheziidae					
<i>Orthezia</i> sp.	Italy	EXT, L	C		5
Pseudococcidae					
<i>Phenacoccus</i> sp.	Italy	EXT, R	C		5
<i>Planococcus</i> sp., prob. <i>citri</i> (Risso)	Italy, Spain	EXT, R	POLY, CP		57
<i>Pseudococcus comstocki</i> (Kuwana)	Canada	EXT, L	CONVOL		13
<i>Pseudococcus longispinus</i> (Targioni-Tozzetti)	Spain	EXT, R	POLY, CP		57
<i>Rhizoecus falcifer</i> Künckel d'Herculis	–	–	POLY		13
HYMENOPTERA					
Formicidae					
<i>Acromyrmex octospinosus</i> (Reich)	Canada	EXT, L	IB		58
Tenthredinidae					
<i>Ametastegia glabrata</i> (Fallén)	USA, Canada	–	CONVOL		1, 13
COLEOPTERA					
Scarabaeidae					
<i>Chasmatopterus</i> sp.	Portugal	EXT, F	C		5
<i>Maladera matrida</i> Argaman	Israel	–	IB		59
<i>Oxythyrea funesta</i> (Poda)	Spain	EXT, F	POLY, CP		60
<i>Phyllophaga ephilda</i> (Say)	USA	EXT, R	IB		61
<i>Tropinota hirta</i> Poda [<i>Epicometis hirta</i>]	Spain	EXT, F	POLY, CP		60
Buprestidae					
<i>Trachys puncticollis</i> Abeille var. <i>obscurella</i> Obenberger	Europe	EXT, L; INT, L	CC	*	5, 62, 63
<i>Trachys puncticollis</i> ssp. <i>rectilineata</i> Abeille de Perin	Slovakia, Austria	EXT, L; INT, L	CA	*	16, 64
<i>Trachys</i> spp.	Uzbekistan Tajikistan	INT, L	CA		21
Elateridae					
<i>Adrastus limbatus</i> (F.)	Yugoslavia	EXT, L	C		5, 65
<i>Conoderus amplicollis</i> (Gyllenhal)	USA	–	IB		66
<i>Conoderus falli</i> Lane	USA	EXT, R	IB		67

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Conoderus rudis</i> Brown	USA	EXT, R	IB		68
<i>Conoderus scissus</i> (Schaeffer)	USA	–	IB		66
<i>Conoderus vespertinus</i> (F.)	USA	EXT, R	IB		67
<i>Conoderus</i> spp.	USA	–	IB		69
Dermestidae					
<i>Attagenus scalaris</i> (Pic)	Egypt	EXT, F	CA		15, 70
Dasytidae					
<i>Astylus variegatus</i> (Germar)	Brazil	–	IB		71
Nitidulidae					
<i>Conotelus obscurus</i> Erichson	–	–	POLY		13
<i>Meligethes solidus</i> (Kugelann)	Spain	EXT, F	POLY		72
<i>Meligethes</i> spp.	Austria, Italy	EXT, F	C		5
Phalacridae					
<i>Olibrus</i> spp.	Italy	EXT, F	CC		5
<i>Stilbus</i> sp.	Italy	EXT, F	CC		5
Coccinellidae					
<i>Subcoccinella vigintiquatuorpunctata</i> (L.)	Italy	EXT, L	POLY, CP		60
Meloidea					
<i>Epicauta ferruginea</i> Say	USA, Canada	EXT, L	POLY		1, 13
<i>Epicauta leopardina</i> Haag	Paraguay	–	I		73
<i>Epicauta trichrus</i> Pallas	USA, Canada	EXT, L	POLY		1, 13
<i>Epicauta vittata</i> (F.)	USA, Canada	EXT, L	POLY		1, 13
<i>Macrobasis immaculata</i> Say	–	–	POLY		13
<i>Macrobasis unicolor</i> (Kirby) [<i>Epicauta murina</i> (LeConte)]	–	–	POLY		13
Lagriidae					
<i>Lagria hirta</i> (L.)	Italy	EXT, L	C		5
<i>Lagria villosa</i> F.	Brazil	–	IB		71
Tenebrionidae					
<i>Gonocephalum pusillum</i> (F.)	Italy	EXT, L	POLY, CP		60
<i>Stenosis angusticollis</i> (Reiche)	Italy	EXT, L	POLY, CP		5
Cerambycidae					
<i>Nupserha apicalis</i> (Fairmaire) [<i>Nupserha apicata</i>]	Africa	INT, St	C	*	74
<i>Nupserha quadrioculata</i> Thunberg	South East Asia	INT, St	C	*	75
Chrysomelidae					
<i>Agrioconota bivittata</i> (Say)	North America	–	C, I		76, 77
<i>Altica oleracea</i> (L.) or nr.	Italy, Spain	EXT, L	POLY, CP		60
<i>Aspidomorpha difformis</i> (Motschulsky)	Japan	–	CJ		13
<i>Aspidomorpha furcata</i> (Thunberg)	Burma, India, Indonesia	EXT, L	IB, IT		78
<i>Aspidomorpha indica</i> Boheman	South East Asia, Pakistan	EXT, L	C, I	*	19, 79
<i>Aspidomorpha miliaris</i> (F.)	South East Asia	–	C, I	*	79
<i>Aspidomorpha transparipennis</i> (Motschulsky)	South East Asia, USA	EXT, L	CONVOL	*	80, 81
<i>Aspidomorpha</i> sp.	Papua New Guinea	–	IB		41

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Aulacophora calva</i> Anand & Cox	India	–	IB		82
<i>Calligrapha praeceles</i> Rogers	North America	–	CS, I		83
<i>Cassida atrata</i> F.	Italy	EXT, L	CA, CS, <i>Salvia</i>	*	5, 84
<i>Cassida circumdata</i> Herbst	South East Asia	–	C, I	*	80, 82
<i>Cassida enervis</i> Boheman	Pakistan	EXT, L	C	*	19
<i>Cassida indicola</i> (Duvivier) [<i>Rhytidocassis indicola</i>]	Pakistan, India	EXT, L	CA, CM	*	19, 85
<i>Cassida nebulosa</i> L.	–	EXT, L	POLY		13
<i>Cassida rubiginosa</i> Müller or nr.	Italy, Kazakhstan	EXT, L	POLY, CA	*	20, 60
<i>Cassida sanguinolenta</i> Müller	Egypt	EXT, L	CA		15
<i>Cassida vespertina</i> Boheman	–	EXT, L	POLY		13
<i>Cassida vittata</i> Villers or nr.	Yugoslavia	EXT, L	POLY, CP		60
<i>Cassida wienmanni</i> Chapuis	Italy	EXT, L	C		5
<i>Cassida</i> sp.	Portugal, Spain	EXT, L	C		5
<i>Cassida</i> sp.	Papua New Guinea	EXT, L	IB		41
<i>Chaetocnema apricaria</i> Suffrian	Brazil	–	IB		71
<i>Chaetocnema basalis</i> Baly	Philippines	EXT, L	IB		34
<i>Chaetocnema confinis</i> Crotch	USA	EXT, R	IB		67, 69
<i>Chaetocnema pulicaria</i> (Melsheimer)	–	–	POLY		13
<i>Charidotella sexpunctata</i> (F.)	Puerto Rico	EXT, L	IB		86
<i>Chelymorpha cassidea</i> (F.)	USA, Canada	EXT, L	CONVOL	*	17, 87–90
<i>Chirida guttata</i> (Olivier) [<i>Deloyala guttata</i>]	North and Central America	EXT, L	CONVOL	*	17, 77, 91
<i>Chrysolina banksi</i> (F.) or nr.	Yugoslavia	EXT, L	POLY, CP		60
<i>Colasposoma sellatum</i> Baly	Australia, Papua New Guinea	EXT, L, St, R	IB		92
<i>Colasposoma subopacum</i> Jacoby	India	–	IB		82
<i>Coptocephala chalybea</i> Germar or nr. [<i>Stolas chalybea</i>]	Portugal	EXT, L	C		5
<i>Coptocyclus aurichalcea</i> F.	North America	–	C		76
<i>Cryptocephalus</i> sp.	Italy	EXT, L	C		5
<i>Deloyala guttata</i> (Olivier)	USA, Canada	EXT, L	CA		1, 90
<i>Diabrotica balteata</i> LeConte	USA, Taiwan	EXT, R	IB		67, 93, 94
<i>Diabrotica duodecimpunctata</i> F.	–	–	POLY		13
<i>Diabrotica speciosa</i> (Germar)	Brazil	–	IB		71
<i>Diabrotica undecimpunctata</i> ssp <i>howardi</i> Barber	USA	EXT, R	IB		67
<i>Diabrotica</i> spp.	USA	–	IB		69, 95
<i>Epitrix parvula</i> (F.) [<i>E. fasciata</i> Blatchley]	–	–	POLY		13
<i>Exosoma lusitanicum</i> (L.) [<i>Exora lusitanica</i>]	Italy, Spain	EXT, F	POLY		96
<i>Galeruca rufa</i> Germar	South and Central Europe, Southern Russia and Asia Minor	EXT, L	CA, CS	*	97, 98
<i>Gastrophysa polygoni</i> (L.)	Italy, Spain	EXT, L	POLY, CP		60
<i>Glyphocassis trilineata</i> (Hope)	Pakistan	EXT, L	CA	*	19
<i>Hypocassida meridionalis</i> (Suffrian)	Italy, Morocco	EXT, L	CC	*	96

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Hypocassida subferruginea</i> Schrank	South and Central Europe, Turkey, Kazakhstan, Uzbekistan, Tajikistan	EXT, L	CC	*	5, 20, 21, 84, 96, 99
<i>Jonthonota nigripes</i> (Olivier)	North America	–	CONVOL		76, 77
<i>Laccoptera quadrimaculata</i> (Thunberg)	South East Asia	–	C, I		80, 82
<i>Leptinotarsa decemlineata</i> Say	North America	EXT, L	POLY		1
<i>Longitarsus aeruginosus</i> (Foudras)	France	–	POLY		100
<i>Longitarsus codinai</i> Madar & Madar	France	EXT, L	C		100
<i>Longitarsus longipennis</i> Kutschera	From Central and East Europe to Central Asia	EXT, L; INT, R	POLY, CONVOL		16, 60, 100, 101
<i>Longitarsus nasturtii</i> (F.)	Portugal, Italy	EXT, L	POLY, CP		60
<i>Longitarsus nigrocillus</i> Motschulsky	Mediterranean region	EXT, L	CONVOL		100
<i>Longitarsus pellucidus</i> (Foudras)	Europe, North Africa, Central Asia, Mongolia, India	EXT, L; INT, R	POLY, CONVOL	*	5, 16, 21, 84, 96, 100, 102–112
<i>Longitarsus rubiginosus</i> (Foudras)	Europe, North Africa, Iran, Caucasus, Siberia	EXT, L; INT, R	CA, CS, <i>Calystegia tricolor</i> , CL		84, 100, 103–105, 110, 113–117
<i>Metriona australica</i> (Boheman) [<i>Cassida australica</i>]	Pakistan, India	EXT, L	CONVOL	*	19
<i>Metriona bicolor</i> (F.) [<i>Charidotella sexpunctata</i>]	North America	EXT, L	CONVOL		1, 17, 88, 90
<i>Metriona purpurata</i> (Boheman) [<i>Charidotella purpurata</i>]	North America	EXT, L	CA		1, 118
<i>Metriona tuberculata</i> (F.) [<i>Charidotella tuberculata</i>]	USA	–	CA, IB		81
<i>Oulema melanopus</i> (L.)	Italy	EXT, L	POLY, CP		60
<i>Phyllotreta consobrina</i> (Curtis)	Europe, North Africa	EXT, L	POLY, CP		60, 100
<i>Physonota jamaicensis</i> (L.) [<i>Eurypepla jamaicensis</i>]	Puerto Rico	–	IB		86
<i>Podagrica malvae</i> (Illiger) or nr.	Europe	EXT, L	POLY, CP		60, 100
<i>Psylliodes glabra</i> L.Redtenbacher or nr.	Italy	EXT, L	C		5
<i>Psylliodes punctulata</i> Melsheimer	–	–	POLY		13
<i>Psyllioides</i> sp.	Italy	EXT, L	C		5
<i>Sphaeroderma testaceum</i> (F.)	Spain	EXT, L	POLY, CP		60
<i>Smaragdina</i> sp.	Portugal	EXT, L	C		5
<i>Strobiderus</i> sp.	India	–	IB		82
<i>Systema blanda</i> (Melsheimer)	North America	EXT, R	IB		67, 119
<i>Systema elongata</i> (F.)	USA	EXT, R	IB		67
<i>Systema frontalis</i> (F.)	USA	EXT, R	IB		67
<i>Systema</i> sp.	Brazil	–	IB		71
<i>Systema</i> sp.	USA	–	IB		69, 95
<i>Typophorus nigrinus</i> (F.)	Brazil	–	IB		71
<i>Typophorus viridicyaneus</i> (Crotch)	–	–	POLY		13
Bruchidae					
<i>Megacerus discoidus</i> Say	USA, Canada	EXT, F; INT, S	CS, IP, ITR	*	17, 90, 120–124
<i>Megacerus impiger</i> (Horn)	USA	INT, S	CC	*	125
<i>Spermophagus calystegiae</i> (Lukjanovič & Ter-Minasjan)	Europe, Turkey, Iran	EXT, F; INT, S	CS	*	84, 126–128

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Spermophagus kuesteri</i> (küsteri) Schilsky	Europe, Turkey, Iran, Israel, Syria, North Africa, Mongolia	EXT, F; INT, S	C	*	84, 126, 127, 129–131
<i>Spermophagus rufiventris</i> Boheman	Japan	EXT, F; INT, S	CJ	*	132
<i>Spermophagus sericeus</i> (Geoffroy)	Europe, North Africa, Iran, Iraq, Turkey, Mongolia, Uzbekistan, Tajikistan	EXT, F; INT, S	CC	*	21, 84, 126, 127, 129–135
<i>Spermophagus variolosopunctatus</i> Gyllenhal	East and South Europe, South Russia, Turkey, North Africa.	EXT, F; INT, S	C	*	48, 126, 127, 136
<i>Spermophagus</i> sp.	Italy, Spain	EXT, F; INT, S	C		5
Curculionidae					
<i>Alcidodes chadoiri</i> (Chevrolat) [<i>A. karelinii</i> (Boheman)]	Uzbekistan, Tajikistan	EXT, S	CA	*	21
<i>Alcidodes fabricii</i> (F.)	Pakistan	EXT, S	CA, I	*	19, 137
<i>Apion basicorne</i> Illiger	Yugoslavia	EXT, L	POLY		138
<i>Apion pisi</i> (F.)	Italy	EXT, L	POLY, CP		60
<i>Apion</i> sp.	Italy	EXT, L	C		5
<i>Calomycterus setarius</i> Roelofs	–	–	POLY		13
<i>Ceutorhynchidius</i> [<i>Ceutorhynchus</i>] <i>baldensis</i> Schultze [<i>Trichosirocalus baldensis</i>]	Italy	EXT, L	POLY		139
<i>Ceutorhynchidius</i> [<i>Ceutorhynchus</i>] <i>maculaalba</i> (Herbst) [<i>Neoglocianus maculaalba</i>]	Italy	EXT, L	POLY		60
<i>Cylas formicarius</i> F.	India, Japan, Taiwan, Kenya, South Africa, Papua New Guinea	EXT, R	IB		27, 82, 140–143
<i>Cylas formicarius</i> ssp. <i>elegantulus</i> (Summers)	Japan, Australia	EXT, R	IB		144, 145
<i>Cylas puncticollis</i> Boheman	South Africa, Nigeria	EXT, R	IB		27, 146
<i>Diaprepes abbreviatus</i> L.	USA, Caribbean islands	–	POLY, IB, CP		147
<i>Euscepes postfasciatus</i> (Fairmaire)	Japan	INT, St, R	IB		148–150
<i>Hypera postica</i> (Gyllenhal)	Italy	EXT, L	POLY		60
<i>Limnobaris pilistriata</i> (Stephens)	Italy	EXT, L	POLY		139
<i>Lixus iridis</i> Olivier	Italy	EXT, L	POLY, CP		60
<i>Otiorynchus</i> sp.	Italy	EXT, L	C		5
<i>Pachnaeus litus</i> (Germar)	Cuba	INT, R	IB		151
<i>Pachytychius squamosus</i> (Gyllenhal)	Italy	EXT, L	POLY, CP		60
<i>Sitona limosus</i> (Rossi)	Italy	EXT, L	POLY, CP		60
<i>Sitona lineatus</i> (L.)	Portugal	EXT, L	POLY, CP		60
<i>Strophomorphus</i> sp. nr. <i>porcellus</i> (Schönherr)	Italy	EXT, L	C		5
<i>Tanymecus palliatus</i> (F.)	Italy	EXT, L	POLY, CP		13, 16, 60
LEPIDOPTERA					
Nepticulidae					
<i>Stigmella freyella</i> (Heyden)	Europe	INT, L	CA, CAL, CS	*	5, 16, 52, 152–157
<i>Stigmella</i> sp.	Austria, Italy, France, Spain, Portugal	INT, L	CC		5
Psychidae					
<i>Apterona helicinella</i> (Herrich-Schäffer)	Europe	INT, L	POLY		154

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Dissoctena granigerella</i> Staudinger	Portugal, Spain	EXT, F	POLY, C		158, 159
Psychidae sp.	South Korea	EXT, L	IB		160
Bucculatricidae					
<i>Bucculatrix cantabricella</i> Chrétien	Slovakia, France, Spain, Italy, Hungary, Yugoslavia	INT, L	<i>Convolvulus cantabricus</i>		154, 157, 161
Bedelliidae					
<i>Bedellia somnulentella</i> (Zeller)	North America, Eurasia, Africa, Australia, Papua New Guinea	INT, L	CONVOL, CP	*	15, 17, 20, 41, 52, 88, 144, 152, 155, 162–165
Ethmiidae					
<i>Ethmia candidella</i> (Alpheraky)	South America, China	–	CA, I		166, 167
Scythrididae					
<i>Scythris lempkei</i> (Bengtsson & Langohr)	France, Spain	EXT, L	POLY		168
<i>Scythris</i> sp.	Italy	EXT, L	C		5
Cosmopterigidae					
<i>Hodgesiella rebeli</i> Krone [<i>Stagmatophora rebeli</i>]	Europe	INT, L	C		152, 154
<i>Pebops</i> sp.	Peru	–	IB		169
Gelechiidae					
<i>Brachmia convolvuli</i> (Walsingham) [<i>Helcystogramma convolvuli</i>]	Philippines	EXT, L	IB		34
<i>Brachmia triannulella</i> ssp. <i>macroscopa</i> (Meyrick) [<i>Helcystogramma triannulella</i> ssp. <i>macroscopum</i>]	South Korea	EXT, L	IB		160
<i>Brachmia</i> sp.	Italy	EXT, L	C		5
<i>Brachmia</i> sp.	Philippines	EXT, L	IB		170
<i>Dichomeris lamprostoma</i> (Zeller)	Spain	INT, L	C		5, 14, 171
<i>Gelechia rhenanella</i> Heyden	Europe	EXT, L	CS		52
<i>Helcystogramma triannulella</i> (Herrich-Schäffer)	Europe, Japan	EXT, L	CONVOL	*	5, 16, 52, 153, 157, 172, 173
<i>Pleurota</i> sp.	Italy	EXT, L	C		5
<i>Trichotaphe</i> sp.	Peru	–	IB		169
<i>Xystophora rhenanella</i> Heyden	Europe	–	C		153, 174
Sesiidae					
<i>Tinthia brosisformis</i> (Hübner)	Russia, East Europe, Turkey	INT, R	C	*	157, 175
<i>Tinthia tineiformis</i> (Esper)	South Europe, Turkey	INT, R	C	*	157, 175
Tortricidae					
<i>Archips crataeganus</i> (Hübner)	Austria	EXT, L	POLY		176
<i>Archips purpuranus</i> Clemens	Canada	EXT, L	CONVOL		13
<i>Argyroploce</i> sp.nr. <i>arbutella</i> (L.)	Austria	EXT, L	CONVOL		5
<i>Argyrotaenia ljugiana</i> (Thunberg)	Italy	EXT, L	CONVOL		177
<i>Cacoecimorpha pronubana</i> (Hübner)	Italy, Spain	EXT, L	POLY		176
<i>Celypha rivulana</i> (Scopoli)	Italy	EXT, L	POLY		172
<i>Cnephasia incertana</i> (Treitschke)	Europe	INT, L	POLY		152, 154, 176, 178, 179,
<i>Cnephasia longana</i> (Haworth)	–	–	POLY		13
<i>Epinotia</i> sp.	Canada	EXT, L	CONVOL		13

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Sparganothis pilleriana</i> (Denis & Schiffermüller)	Austria	EXT, L	POLY, CP		176
Pterophoridae					
<i>Emmelina monodactyla</i> (L.)	Europe, Africa, Asia, North and Central America	EXT, L	POLY, CONVOL	*	1, 5, 15–17, 88, 152, 155, 180–191
<i>Ochyrotica fasciata</i> Walsingham	Peru	–	IB		169
<i>Pterophorus ischnodactyla</i> (Treitschke)	South Europe	EXT, L	<i>Convolvulus cantabricus</i>		157, 191
<i>Pterophorus jezonicus</i> Matsumura	Asia	–	C, I		192
<i>Pterophorus pentadactyla</i> (L.)	Europe	EXT, L	POLY	*	5, 16, 152, 191, 193–195
<i>Pterophorus</i> sp.	South East Asia	–	C		196
<i>Stenoptilia pterodactyla</i> (L.)	Europe	EXT, L	POLY		52, 152, 159, 195
Pyralidae					
<i>Aporodes floralis</i> (Hübner) [<i>Noctuella floralis</i>]	Syria, Afghanistan, Egypt, India, Europe	EXT, L, St	CC	*	5, 15, 16, 18, 157, 197
<i>Chrysocrambus linetellus</i> (F.)	Europe	EXT, L	CONVOL	*	5, 157
<i>Diastichtis</i> [<i>Anomostictis</i>] <i>fracturalis</i> Zeller	North America	–	C		88
<i>Eurrhypara hortulata</i> (L.)	Europe	EXT, L	CA, CS		52
<i>Hydrisis ornatalis</i> (Duponchel)	Switzerland, South Europe, Egypt	EXT, L	CONVOL	*	5, 15, 157, 159
<i>Loxostege mancalis</i> (Lederer) [<i>Hahncappisia mancalis</i>]	USA, Canada	–	CA		1
<i>Loxostege sticticalis</i> (L.)	Canada, USA, Italy, Yugoslavia	EXT, L	POLY, CP		1, 198, 199
<i>Macrobotys pertextalis</i> (Lederer) [<i>Herpetogramma pertextalis</i>]	Canada	–	CONVOL		13
<i>Microthyris anormalis</i> (Guenée)	Peru	EXT, L	IB		169, 200
<i>Omphisa anastomosalis</i> Guenée	Asia, Papua New Guinea, Taiwan	INT, St, R	IB		41, 201, 202
<i>Phlyctaenia coronata</i> (Hufnagel)	Europe	–	POLY		52, 152
<i>Pyrausta despicata</i> (Scopoli)	Italy, Spain	EXT, L	POLY		194, 193, 203
<i>Sitochroa verticalis</i> (L.)	South Europe	EXT, L	POLY		193
<i>Titanio normalis</i> (Hübner)	Eurasia, Africa	INT, L	CC	*	16, 154, 157
Sphingidae					
<i>Coelonia fulvinotata</i> (Butler)	–	–	POLY		13
<i>Deilephila elpenor</i> (L.)	Eurasia, Africa	EXT, L	CS		157, 204, 205
<i>Herse convolvuli</i> L. [<i>Agrius convolvuli</i>]	Europe, India, China, Oman, Egypt, Saudi Arabia, Madagascar, South Africa, Japan, Australia, Papua New Guinea, Hawaii	EXT, L	C, POLY, CP		15, 16, 21, 41, 144, 199, 206–216
<i>Hippotion celerio</i> (L.)	Europe	EXT, L	POLY, IB		144, 157
<i>Hippotion scrofa</i> (Boisduval)	Australia	–	IB		144
<i>Theretra nessus</i> (Drury)	South East Asia	–	C		217
Geometridae					
<i>Ascotis selenaria</i> (Denis & Schiffermüller)	Europe, Israel, Japan	EXT, L	POLY, CP		48, 152, 218–222
<i>Ematurga atomaria</i> (L.)	Italy	EXT, L	POLY, CP		48, 152, 207

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Idaea degeneraria</i> (Hübner)	–	EXT, L	POLY		211
<i>Idaea emarginata</i> (L.)	Europe	EXT, L	POLY		52, 211
<i>Idaea</i> sp.	Italy	EXT, L	C		5
<i>Larentia clavaria</i> (Haworth)	Italy	EXT, L	CC, Malvaceae		5, 206
<i>Lycia florentina</i> (Stephanelli)	Italy, Yugoslavia	EXT, L	POLY, CP		48, 157
<i>Orthonama obstipata</i> (F.)	–	–	POLY		223
<i>Scopula emissaria</i> (Walker)	Korea	EXT, L	IB		160
<i>Scopula emutaria</i> Hübner	Europe	EXT, L	POLY		152
<i>Scopula rubiginata</i> (Hufnagel)	Europe	EXT, L	POLY		152, 223
<i>Semiothisa clathrata</i> (L.) [<i>Chiasmia clathrata</i>]	Italy	EXT, L	LEG, CA		5, 206
Notodontidae					
<i>Schizura ipomoeae</i> Doubleday	–	–	POLY		13
Noctuidae					
<i>Acontia lucida</i> Hufnagel	Europe, Asia	EXT, L	POLY		152, 211, 224
<i>Acronicta rumicis</i> (L.)	Italy	EXT, L	POLY		193
<i>Aedia funesta</i> (Esper)	Europe, Turkey	EXT, L	CC	*	16, 152, 157, 225, 226
<i>Aedia leucomelas</i> (L.)	Europe, Turkey, Japan, Korea	EXT, L	CONVOL		52, 157, 160, 172, 173
<i>Agrotis ipsilon</i> (Hufnagel)	Egypt, Italy	EXT, L	POLY, CP		15, 199
<i>Agrotis segetum</i> Dennis & Schiffermüller	Italy	EXT, L	POLY, CP		38
<i>Anepischetos</i> [<i>Hypena</i>] <i>lividalis</i> (Hübner)	Europe	–	C, <i>Parietaria</i>		5, 157, 227
<i>Autographa gamma</i> (L.)	Worldwide	EXT, L	POLY, CP		193, 228
<i>Axylia putris</i> (L.)	Italy, Poland, Russia	EXT, L	POLY		21, 152, 199, 206, 229
<i>Chrysodeixis chalcites</i> Esper	Italy	EXT, L	POLY, CP		230
<i>Discestra trifolii</i> (Hufnagel)	Italy	EXT, L	POLY		48
<i>Elaphria morpheus</i> Hufnagel [<i>Caradrina morpheus</i>]	–	–	POLY		13
<i>Emmelia trabealis</i> (Scopoli)	Europe, Asia	EXT, L	CA, CS	*	5, 16, 21, 52, 152, 193, 211, 224, 226
<i>Estigmene acraea</i> (Drury)	–	–	POLY		13
<i>Eublemma baccalix</i> (Swinhoe)	Pakistan	EXT, F	CA, I		18, 137
<i>Euplexia lucipara</i> L.	–	–	POLY		13
<i>Euxoa segetum</i> Denis & Schiffermüller [<i>Agrotis segetum</i>]	–	–	POLY		13
<i>Heliothis armigera</i> (Hübner) [<i>Helicoverpa armigera</i>]	Italy, Sweden, Canada, USA, India, China, Egypt	EXT, L	POLY, CP		1, 231–234
<i>Lacanobia oleracea</i> (L.)	Italy, Spain, UK, Bulgaria, Slovakia	EXT, L	POLY, CP		193, 235–237
<i>Lycophotia margaritosa</i> (Haworth) [<i>Peridroma saucia</i> Hübner]	USA, Canada	–	CONVOL		1
<i>Mamestra brassicae</i> (L.)	Italy, Bulgaria, Yugoslavia, France, Slovakia	EXT, L	POLY, CP		193, 198, 199, 237–239
<i>Melanchra pisi</i> (L.)	Europe	EXT, L	POLY		52
<i>Mythimna</i> sp.	Spain	EXT, L	C		5

Table 1. Organisms associated with the family Convolvulaceae worldwide^a.

Taxon ^b	Location	Plant association ^c	Host specificity ^d	Biocontrol candidate ^e	Ref.
<i>Neleucania diffusa</i> (Walker) [<i>Faronta diffusa</i>]	–	–	POLY		13
<i>Peridroma saucia</i> (Hübner)	Canada, Italy	EXT, L	POLY, CP		13, 39, 199, 240
<i>Pseudaletia unipuncta</i> (Haworth) [<i>Mythimna unipuncta</i>]	Canada, USA	–	CONVOL		13
<i>Rivula sericealis</i> Scopoli	France	EXT, L	POLY		206
<i>Spodoptera eridania</i> Cramer	Brazil, Peru	–	POLY, IB		241, 242
<i>Spodoptera exigua</i> (Hübner)	Europe, Egypt, USA, Canada	EXT, L	POLY, CP		1, 39, 211
<i>Spodoptera littoralis</i> Boisduval	Italy, Egypt	EXT, L	POLY		15
<i>Spodoptera ornithogalli</i> (Guenée)	–	–	POLY		13
<i>Synthymia fixa</i> (F.)	Europe	EXT, L	C, <i>Psoralea</i> ,		5, 157, 227
<i>Trachea atriplicis</i> (L.)	Europe	–	CS, C		243
<i>Tyta luctuosa</i> (Denis & Schiffermüller)	Europe, Asia, North Africa	EXT, L	CC	*	16, 21, 33, 152, 157, 224 – 226, 244
<i>Xylena vetusta</i> (Hübner)	Austria	EXT, L	POLY		193
Arctiidae					
<i>Arctia caja</i> L.	–	–	POLY		13
<i>Diacrisia virginica</i> (F.) [<i>Spilosoma virginica</i>]	USA, Canada	–	POLY		1, 13
<i>Eilema</i> sp.	Italy	EXT, L	CS		5
DIPTERA					
Cecidomyiidae					
<i>Lasioptera convolvuli</i> Felt	North America	INT, St	CA, CS	*	13, 245, 246
Agromyzidae					
<i>Melanagromyza albocilia</i> Hendel	Slovakia, Austria, Hungary, UK, Italy, Israel, Egypt	INT, St, R	CA	*	5, 15, 247 – 249
<i>Melanagromyza convolvuli</i> Spencer	Pakistan	INT, St	CA	*	18, 247, 250
<i>Chromatomyia horticola</i> (Goureaux)	Worldwide	INT, L	POLY, CP		154

^aFamilies are listed taxonomically, and genera and species within families in alphabetical order. ^bTaxa are largely listed as cited in the references. Where appropriate, preferred synonyms are given in square brackets. ^cEXT: external feeding; INT: internal feeding; L: leaf; St: stem; R: root; S: seed; SC: seed capsule. ^dCA: *Convolvulus arvensis*; CAL: *Convolvulus althaeoides*; CJ: *Calystegia japonica*; CL: *Convolvulus lanceolata*; CM: *Convolvulus microphylus*; CS: *Calystegia sepium*; C: *Convolvulus* spp.; CC: species of *Convolvulus* and *Calystegia*; CONVOL: members of the Convolvulaceae; Convall.: *Convallaria*; I: *Ipomoea* sp.; IB: *Ipomoea batatas* (sweet potato); ID: *Ipomoea denticulata*; IP: *Ipomoea pandurata*; IS: *Ipomoea staphilina*; IT: *I. tuberosa*; ITR: *Ipomoea tricolor*; LEG: Leguminosae; LB: *Lepistemon binctarifus*; MG: *Merremia gemella*; POLY: polyphagous species; CP: crop pest. ^eBiological control candidate according to the literature.

3.1. Gall-Forming Mite

Feeding by *Aceria malherbae* Nuzzaci (Acari, Eriophyidae) on young leaf petioles, stems and expanding buds causes abnormal growth or galling. Flowers have never been seen on stems heavily infested with the mites [29, 32]. The symptoms were found only in natural ecosystems and were never recorded in cultivated fields [16]. The mite lived and reproduced only on species in the genera *Convolvulus* and *Calystegia* [31]. In 1989, *A. malherbae* was released in Saskatchewan (Canada) and New Jersey and Texas (USA) where it established [22]. During 1993–98, *A. malherbae* was released and established in southern Alberta (Canada) and Montana (USA). Populations of the mite survived for up to 4 years at some localities and caused slight to severe damage to the weed.

The establishment of *A. malherbae* in these areas suggests that it could establish throughout the North American range of *Convolvulus arvensis* [252].

3.2. Root and Stem Feeders

Melanagromyza albocilia Hendel (Dipt., Agromyzidae), a native European stem borer of *C. arvensis*, was considered as a very promising candidate for the biological control of bindweed [5, 249]. Larvae bore tunnels in bindweed stems and root crowns, causing the death of infested shoots [247]. The number of infested plants ranged from 46.7% to 99.2% and the number of infested stems from 4.1% to 37.2% in southwest Slovakia [253]. The fly can promote secondary damage due to infection by pathogenic microorganisms [254]. Its feeding habit in stems indicates a high degree of probability that the species

Table 2. Systematic affiliation of organisms associated with the family Convolvulaceae worldwide.

Order	%	Number		
		Species	Families	Biological control candidate species
Tylenchata	1.8	6	3	0
Acari	3.1	10	2	1
Collembola	0.3	1	1	0
Orthoptera	0.9	3	2	0
Thysanoptera	0.9	3	1	0
Hemiptera	13.6	45	13	0
Hymenoptera	0.6	2	2	0
Coleoptera	43.7	143	15	29
Lepidoptera	33.9	111	17	14
Diptera	1.2	4	2	3
Total	100.0	328	58	47

Table 3. Feeding sites of organisms found on the family Convolvulaceae worldwide^a.

External or internal feeding on:	Number of species	%
Leaves	177	54.0
Roots	26	8.0
Seeds, seed capsules	16	4.9
Flowers	11	3.3
Stems	11	3.3
Not specified	87	26.5
Total	328	100.0

^aIf an insect fed on different part of the plants as an adult and as a larva, it is included here in its more destructive stage.

will prove to be strictly monophagous [255]. Its known distribution includes Austria and Hungary [256, 257], Egypt [15], Israel [258], UK, Denmark and Sweden [248], Italy [5] and the Czech Republic [259]. High numbers of *M. albocilia* were recorded only in Slovakia [16].

Melanagromyza convolvuli Spencer was found in *C. arvensis* stems in Pakistan [250]. Larval feeding in the stems or lateral branches considerably weakened the plants. Infested branches dried up under the combined effects of drought and larval attack in areas with limited rainfall [247]. However, the species was able to complete its development on sweet potato and *Ipomoea purpurea* (L.) Roth [19]. For this reason *M. convolvuli* can be used as biological control agent only in areas where sweet potato is not grown as a crop [7].

The adult of the *Convolvulus* flea beetle, *Longitarsus pellucidus* (Foudras) (Col., Chrysomelidae), was found to be a widespread and abundant leaf feeder. The larva fed on roots. Gorenshstein [21] mentioned

L. pellucidus as highly specialized on *C. arvensis*. In Italy, the adults were found primarily on *Convolvulus* species. Only slight feeding on sweet potato was observed. This species seems to have the most potential as a classical biological control agent for introduction from Mediterranean Europe [5]. *Longitarsus pellucidus* is well synchronized with the host plant and if released as a biological control agent in the classical concept, its population should be able to increase to a size large enough to suppress *C. arvensis*. *Longitarsus pellucidus* is widely distributed in the Old World, from the British Isles to Mongolia and India. It has also been reported from North Africa and the Middle East (Table 1). LeSage [260] reported the species for the first time in North America.

Nupserha apicata (Fairmaire) (syn. *N. apicalis*) (Col., Cerambycidae) in Africa [74] and *Nupserha quadrioculata* Thunberg in southeastern Asia [75] feed in stems of *Convolvulus* spp. forming galls [74]. The species are able to suppress weed populations.

3.3. Seed Feeders

Species from the family Bruchidae (Coleoptera) were frequently associated with *C. arvensis* and *Calystegia sepium*. In the Palaearctic region, *Spermophagus sericeus* (Geoffroy), *Spermophagus calystegiae* (Lukjanoviæ & Ter-Minasjan., *Spermophagus kuesteri* Schilsky and *Spermophagus variolosopunctatus* Gyllenhal [127] and *Spermophagus rufiventris* Boheman [132] were reared from *Convolvulus arvensis* seeds. *Megacerus discooides* Say [123] and *Megacerus impiger* (Horn) [125] were reported as native North American seed feeders on *Calystegia sepium*.

In Europe, the most common bruchid species is *S. sericeus*. Adults feed on pollen of different plant species mainly in the Convolvulaceae and larvae feed in seeds [134]. In some cases *S. sericeus* has been able to generate high levels of infestation even after only one generation on the host. More than 50% of *Convolvulus arvensis* seeds were attacked by this insect in Slovakia [16]. In Italy, infestation levels of 40% were found at Rome and 65% at Venice [5]. *Megacerus discooides* destroyed about 63% of the viable seeds of *Calystegia sepium* at Belleville in Canada [17]. However, the species is not always able to generate high levels of infestation, probably because of parasitism. The egg stage is particularly vulnerable because eggs are laid on the surface of the ripening pod. Bridwell [261] noted that when the trichogrammatid egg parasite *Uscana semifumipennis* Girault was present, only six larvae reached maturity out of 3000 eggs of the bruchid *Caryedon serratus* (Olivier). A similar species, *Uscana spermophagi* Viggiani, was found to be a parasitoid of *S. sericeus* [262].

The newly hatched larvae of *Alcidodes fabricii* (F.) (Col., Curculionidae) bored upwards into the fruits and destroyed all seeds of *Convolvulus arvensis* in Pakistan. The species appeared to be specific to *C. arvensis* although it would infest *Ipomoea* spp. [137]. The larvae of a related species, *Alcidodes karelinii*

(Boheman) (syn. *A. chaudiroidi* [Chevrolat]), fed on the reproductive organs of *C. arvensis* in Uzbekistan and Tajikistan [21].

3.4. Foliage Feeders

Galeruca rufa Germar (Col., Chrysomelidae), a European species, feeds only on plant species in the genera *Convolvulus* and *Calystegia* [97]. Moderate populations of this insect can cause defoliation severe enough to reduce *Convolvulus arvensis* flower production and to kill seedlings [98]. Natural populations of *G. rufa* could be expected to occur on *C. arvensis* continuously through the season (with feeding by both adults and larvae) [263]. *Galeruca rufa* was considered by the above authors as a promising biological control agent for *C. arvensis* in the USA.

Tortoise beetles (Col., Chrysomelidae), including *Rhytidocassis indicola* (DuRoi) (syn. *Cassida indicola*) [85], *Aspidomorpha indica* Boheman [19], *Aspidomorpha miliaris* (F.) [79], *Aspidomorpha transparipennis* (Motschulsky) and *Cassida circumdata* Herbst [80] and *Cassida enervis* Boheman, *Glyphocassis trilineata* (Hope) and *Cassida australica* (Boheman) (syn. *Metriorhina australica*) [18] were found to be promising agents in Pakistan and India. Each of them can defoliate *Convolvulus arvensis* and (or) *Calystegia sepium* severely when their populations are high. However, their value in biological control is limited to areas where sweet potato is not grown, because they all feed on this plant.

Cassida rubiginosa Müller was mentioned as a prospective phytophagous species for the biological control of *Convolvulus arvensis* and *Calystegia sepium* in Kazakhstan [20], and *Cassida atrata* F. was suggested for the same purpose in Italy [5].

Hypocassida subferruginea Schrank and *Hypocassida meridionalis* (Suffrian) (Col., Chrysomelidae), were recorded as specific to the genera *Convolvulus* and *Calystegia* in Eurasia (Table 1). The larvae and adults of *H. subferruginea* almost completely destroyed the leaves of *Convolvulus arvensis* plants in some uncultivated habitats in the warmest localities of Slovakia [16]. Feeding tests showed that 28 newly hatched larvae of *H. subferruginea* fed only 6% as much on sweet potato as control larvae did on *C. arvensis*. Only four of them survived the 5-day test, and none of them completed development on sweet potato [5].

Chelymormpha cassidea (F.) (Col., Chrysomelidae), a frequent defoliator of bindweeds in the USA (Table 1), also showed potential for their control. Larvae and adults were successfully established on *Calystegia sepium* and *Convolvulus arvensis* in rye, maize, grapes, pine and zucchini. This insect fed only on plants within the Convolvulaceae and can be reared in the laboratory [264].

The larvae of *Tyta luctuosa* (Denis & Schiffermüller) (Lep., Noctuidae) generally feed during the night on

leaves and flowers of *Convolvulus* or *Calystegia* throughout Europe and Asia (Table 1). The species was released for biological control purposes in the USA (1987) and Canada (1990) but its establishment was not confirmed [22].

On the basis of the results from Slovakia, *Emmelina trabealis* (Scopoli) from the same family is probably more promising than *T. luctuosa*. The larvae of *E. trabealis* fed during the daytime on leaves, but also flowers, where they evidently reduced reproductive capacity in *Convolvulus arvensis* as a result of feeding on pistils and stamens [16]. Larvae fed only lightly and were unable to complete development on sweet potato [5].

In Europe, Syria, Afghanistan, Pakistan, India and Egypt, the larvae of *Noctuelia floralis* Hübner (syn. *Aporodes floralis*) (Lep., Pyralidae) were noted as very promising biological control agents feeding on young leaves and buds, and subsequently the soft, whitish, underground or aerial stems of bindweeds [5, 15, 19, 197].

The plume moth *Emmelina monodactyla* (L.) is a member of the family Pterophoridae, whose larvae are primarily leaf feeders or stem and root borers on a wide range of host plants. First-instar larvae fed gregariously on the leaves and spread out on the plants as they developed [184]. In Slovakia, *E. monodactyla* was selected as a potential agent for biological control because of its distinct preference for *C. arvensis* and high abundance throughout the area [265]. Smith [88], Mohyuddin [17] and Ghani *et al.* [196] also considered its use as a biological control agent of bindweeds. Sweet potato is of North or South American origin [251] and is grown all over the world [266]. *Emmelina monodactyla* is distributed nearly worldwide [191], but only in one case was damage recorded on sweet potato [267]. Generally, sweet potato was free from very serious attack by *E. monodactyla* [120]. However, the moth's ability to complete development on nine sweet potato varieties under laboratory conditions [186] precludes its use as a control agent in the areas where this crop is grown. Nonetheless, Rosenthal & Buckingham [5] recommended *E. monodactyla* for biological control purposes. They considered that larvae could not survive more than 2 weeks on sweet potato and, moreover, they were not able to complete development on this plant. According to Parrella & Kok [268], releases of three and five larvae of *E. monodactyla* per *Calystegia sepium* leaf caused 90% and 100% reduction of foliage in the greenhouse, respectively. A similar test, where *C. sepium* was infested with three larvae of *E. monodactyla* per leaf in field cages, resulted in a decrease in total length of sprouts, and number of leaves or shoots. In this test, average larval leaf consumption was 13 cm² at 18.3°C, increasing to 16 cm² at 29.4°C. The last-instar larvae consumed over 60% of the leaf area [184]. Inundative releases of ten larvae of *E. monodactyla* per *Convolvulus arvensis* plant caused full defoliation of plants under laboratory conditions. Although the population density of *E. monodactyla* under field conditions

was found to be low [16], a well-managed conservation biological control strategy could provide a method of controlling *C. arvensis* in the field [265].

Larvae of *Helcystogramma triannulella* (Herrich-Schäffer) (Lep., Gelechiidae) were commonly found in rolled leaves of *C. arvensis* and *Calystegia sepium* in Europe [16, 52, 153, 172] and Canada [17]. However, this species was recorded as a foliage feeder on sweet potato in Japan, Korea and the Philippines [34, 160, 170, 173]. Hence, on the world scale, *H. triannulella*'s value is limited to areas with no sweet potato production.

Multiple infestation by larvae of *Bedellia somnulentella* (Zeller) (Lep., Bedelliidae) in a single bindweed leaf generally ends with complete destruction of the leaf, since the mines amalgamate into a single large mine that extends throughout the leaf tissue [15]. The accumulation of frass can result in fungal growth causing more harm to the plant [163]. Although, Tawfik *et al.* [163] suggested *B. somnulentella* for biological control purposes in Egypt, this species feeds on sweet potato [164, 269, 270].

4. Conclusions

This review highlights that organisms associated with the family Convolvulaceae have great diversity even within single families, and only a few species have been studied in detail. In particular, tropical faunas are largely unstudied (apart from organisms feeding on sweet potato) and these are almost certainly greater in size than temperate faunas. So it is still questionable how representative our current knowledge is.

Convolvulus arvensis and other species in the Convolvulaceae have more than one means of reproduction, i.e. seeds and roots. Thus complete biological control of this weed is unlikely to be achieved by using a single species which causes damage limited to one plant part, for example seeds or leaves.

Research efforts should be directed towards continued investigation of phytophagous arthropods attacking the Convolvulaceae, and how suitable species could be effectively combined to put maximum stress on *C. arvensis*. Such efforts may lead to an effective integrated pest management programme. The variety of biological control agents available (identified in Table 1) indicates a reasonable probability of success. From results obtained in Slovakia, we have every reason to believe that *Melanagromyza albocilia*, *Longitarsus pellucidus*, *Hypocassida subferruginea*, *Spermophagous sericeus*, *Emmelina monodactyla* and *Emmelia trabealis* are the most promising [16]. These species cause heavy damage to the host plant and their combined influence is evident in each growing season and on each part of *C. arvensis*.

It is clear that insects have considerable success in controlling *C. arvensis* in natural ecosystems, uncultivated areas and vineyards with living green cover.

Given this, as well as public concern about environmental problems caused by modern agriculture, the development of new cropping systems could help in managing *C. arvensis*.

Because sweet potato and some ornamental plants in the genus *Ipomoea* are often subjected to attack by insects associated with *C. arvensis*, conflicting interests will be the major hurdle in the search for biological control agents that can be used against the weed on the world scale. However, by finding stenophagous species to control *C. arvensis*, the benefits of control could be more easily reconciled with the economic and ecological value of sweet potato and ornamental *Ipomoea* species.

5. Acknowledgements

This work was supported by the Science and Technology Assistance Agency of Slovakia under Contract No. APVT-51-022002 and the Grant Agency of Slovak Agricultural University No. 707/023 20.

6. References

- Weaver, S.E. & Riley, W.R. The biology of Canadian weeds. 53. *Convolvulus arvensis* L. *Canadian Journal of Plant Science* **62** (1982) 461–472.
- Westra, P., Chapman, P., Stahlman, P.W., Miller, S.D. & Fay, P.K. Field bindweed (*Convolvulus arvensis*) control with various herbicide combinations. *Weed Technology* **6** (1992) 949–955.
- Klimeš, L.; Klimešová, J. Biomass allocation in a clonal vine: effects of intraspecific competition and nutrient availability. *Folia Geobotanica and Phytotaxonomica* **29** (1994) 237–244.
- Maillet, J. Les liserons. *Phytoma* **399** (1988) 11–15.
- Rosenthal, S.S. & Buckingham, R.G. Natural enemies of *Convolvulus arvensis* in western Mediterranean Europe. *Hilgardia* **5** (1982) 1–19.
- Andres, L.A. Conflicting interests and the biological control of weeds. In *Proceedings, 5th International Symposium on Biological Control of Weeds*, Brisbane, Australia, 22–27 July 1980 (1981) pp. 11–22.
- Wang, R. & Kok, L.T. Bindweeds and their biological control. *Biocontrol News and Information* **6** (1985) 303–310.
- Huffaker, C.B. Fundamentals of biological weed control. In *Biological control of insect pest and weeds*, DeBach, P. (Ed) Reinhold, New York (1964) pp. 631–649.
- Andres, L.A., Davis, C.J., Harris, P. & Wapshere, A.J. Biological control of weeds. In *Theory and practise of biological control*, Huffaker, C.B. & Messenger, P.S. (Eds) Academic Press, New York (1976) 788 pp.
- Pfirter, H.P., Ammon, H.U., Guntli, D., Greaves, M.P. & Défago, G. Towards the management of field bindweed (*Convolvulus arvensis*) and hedge bindweed (*Calystegia sepium*) with fungal pathogens and cover crops. *Integrated Pest Management Reviews* **2** (1997) 61–69.
- Schroeder, D. Biological control of weeds. In *Recent advances in weed research*, Fletcher, W.W. (Ed) Commonwealth Agricultural Bureaux, Slough, UK (1983) 266 pp.

- 12 Schroeder, D. Biological control of weeds: a review of principles and trends. *Pesquisa Agropecuaria Brasileira* **27** (1992) 191–212.
- 13 Mohyuddin, A.I. Insects from *Calystegia* spp. and *Convolvulus* spp. *Technical Bulletin, Commonwealth Institute of Biological Control* **11** (1969) 93–104.
- 14 Baloch, G.M. Phytophagous organisms associated with bindweed, *Convolvulus* spp., in Pakistan. *Technical Bulletin, Commonwealth Institute of Biological Control* **17** (1974) 29–36.
- 15 Awadallah, K.T., Tawfik, M.F.S. & Shalaby, F.F. Insect fauna of bind-weed, *Convolvulus arvensis* L., in Giza, Egypt. *Bulletin de la Société Entomologique d'Égypte* **60** (1976) 15–24.
- 16 Tóth, P. *Insects – a fresh perspective in the biological control of field bindweed* (*Convolvulus arvensis* L.). PhD thesis, Slovak Agricultural University, Nitra, Slovakia (2000) 229 pp.
- 17 Mohyuddin, A.I. The biology and host spectrum of some stenophagous insects found on *Convolvulus* and *Calystegia* spp. at Belleville, Ontario. *Technical Bulletin, Commonwealth Institute of Biological Control* **12** (1969) 131–146.
- 18 Baloch, G.M. Insects as biological control agents of field bindweed, *Convolvulus arvensis*. *PANS* **23** (1977) 58–64.
- 19 Baloch, G.M. Tortoise-beetles (Chrysomelidae: Cassidinae) associated with field-bindweed, *Convolvulus arvensis* L. in Pakistan. *Technical Bulletin, Commonwealth Institute of Biological Control* **18** (1977) 137–144.
- 20 Tyurebaev, S.S. Prospective phytophages for biological control of certain weeds of Eurasian origin. *Izvestiya Akademii Nauk Kazakhskoi SSR, Biologicheskaya* **2** (1981) 32–35.
- 21 Gorenshtein, B.M. A promising weed eater. *Zashchita Rastenii* **7** (1982) 1–30.
- 22 Julien, M.H. (Ed) *Biological control of weeds: a world catalogue of agents and their target weeds*, 3rd ed. CSIRO, Brisbane, Australia (1992) 194 pp.
- 23 Buhr, H. *Bestimmungstabellen der Gallen (Zoo- und Phyto-ccidien) an Pflanzen Mittel- und Nord-Europas*, Vol. I. Gustav Fischer, Jena, German Democratic Republic (1964) 761 pp.
- 24 Sturhan, D. Outdoor occurrence of *Meloidogyne* species in Western Germany. *Nachrichtenblatt des Deutschen Pflanzenschutzdienstes* **28** (1976) 113–117.
- 25 Salem, F.M., Mohamed, A.M., Abu-Elghar, M.R. & Osman, G.Y. Biological studies of root-knot nematodes on certain weeds, ornamentals, vegetables and field crops. *Minufiya Journal of Agricultural Research* **7** (1983) 355–368.
- 26 Salawu, E.O. & Afolabi, S.S. Weed hosts of a root-knot nematode, *Meloidogyne incognita*, at the Bacita Sugarcane Plantation, Nigeria. *Pakistan Journal of Nematology* **12** (1994) 67–71.
- 27 Daiber, K.C. Injurious insects, spider mites and nematodes on sweet potato in southern Africa. *Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz* **101** (1994) 550–557.
- 28 Arjun, L.A.L., Yadav, B.S. & Nandwana, R.P. A record of some new and known weed hosts of *Rotylenchulus reniformis* Linford & Oliveira, 1940 from Rajasthan. *Indian Journal of Nematology* **6** (1976) 94–116.
- 29 Rosenthal, S.S. Current status and potential for biological control of field bindweed, *Convolvulus arvensis*, with *Aceria convolvuli*. In *Biological control of pests by mites. Proceedings of a conference, April 5–7, 1982*, Hoy, M.A., Cunningham, G.L. & Knutson, L. (Eds) University of Agriculture and Natural Resources, CA, USA, Special Publication No. 3304 (1984) pp. 57–60.
- 30 Nuzzaci, G., Mimmochi, T. & Clement, S.L. A new species of *Aceria* (Acari: Eriophyidae) from *Convolvulus arvensis* L. (Convolvulaceae) with notes of other eriophyids associated with convolvulaceous plants. *Entomologica* **20** (1985) 81–89.
- 31 Rosenthal, S.S. & Platts, B.E. Host specificity of *Aceria (Eriophyes) malherbae* (Acari: Eriophyidae), a biological control agent for weed, *Convolvulus arvensis* (Convolvulaceae). *Entomophaga* **35** (1990) 459–463.
- 32 Craemer, C. *Eriophyidae (Acari) as potential control agents of South African weeds, with descriptions of a new species of Tegenotus Nalepa*. M.Sc. thesis, Rand Afrikaans University, Johannesburg, South Africa (1993).
- 33 Rees, N.E. & Rosenthal, S.S. Field bindweed. In *Biological control of weeds in the West*, Rees, N.E., Quimby, P.C., Piper, G.L., Coombs, E.M., Turner, C.E., Spencer, N.R. & Knutson, L.V. (Eds) Western Society of Weed Science, Bozeman, MT, USA (1996) 342 pp.
- 34 Amalin, D.M., Vasquez, E.A. & Vanderzaag, P. Note: arthropods of sweet potato in the Philippines. *Philippine Agriculturist* **74** (1991) 39–50.
- 35 Mohanasundaram, M. Further studies on the eriophyid fauna (Eriophyoidea: Acari) of Tamil Nadu. *Entomon* **16** (1991) 187–192.
- 36 Jeppson, L.R., Keifer, H.H. & Baker, E.W. *Mites injurious to economic plants*. University of California Press, Berkeley, CA, USA (1975) 614 pp.
- 37 Dabrowski, Z.T. & Marczak, Z. Studies on the relationship of *Tetranichus urticae* Koch and host plants. I. Effect of plant species. *Polskie Pismo Entomologiczne* **42** (1972) 821–855.
- 38 Edwards, C.A. & Heath, G.W. *The principles of agricultural entomology*. Chapman & Hall, London (1964) 418 pp.
- 39 Metcalf, C.L. & Flint, W.P. *Destructive and useful insects*, 4th ed. McGraw-Hill, New York (1962) 1087 pp.
- 40 Stichel, W. *Illustrierte Bestimmungstabellen der Wanzen. II. Europa*, Vol. 4. Hermsdorf, Berlin, German Federal Republic (1955–62).
- 41 Bourke, R.M. Sweet potato (*Ipomoea batatas*) production and research in Papua New Guinea. *Papua New Guinea Journal of Agriculture, Forestry and Fisheries* **33** (1985) 89–108.
- 42 Snodgrass, G.L., Scott, W.P. & Smith, J.W. An annotated list of the host plants of *Lygus lineolaris* (Hemiptera: Miridae) in the Arkansas, Louisiana, and Mississippi delta. *Journal of the Georgia Entomological Society* **19** (1984) 93–101.
- 43 Grenčík, M. *Ochrana rastlín proti živočíšným škodcom*. Príroda, Bratislava, Czechoslovakia (1989) 369 pp.
- 44 Maixner, M. Transmission of German grapevine yellows (Vergilbungskrankheit) by the planthopper *Hyalesthes obsoletus* (Auchenorrhyncha: Cixiidae). *Vitis* **33** (1994) 103–104.
- 45 Maixner, M., Ahrens, U. & Seemuller, E. Detection of the German grapevine yellows (Vergilbungskrankheit) MLO in grapevine, alternative hosts and a vector by a specific PCR procedure. *European Journal of Plant Pathology* **101** (1995) 241–250.

- 46 Boudon, P.E. Black wood. Some unknown factors are lifted, but others remain unknown. *Phytoma* **488** (1996) 10–13.
- 47 Edwards, J. *The Hemiptera – Homoptera of British islands*. Reeve & Co., London (1896) 271 pp.
- 48 Grandi, G. *Introduzione allo studio dell'entomologia*, Vol. 2. Edizioni Agricole, Bologna, Italy (1951).
- 49 Bonnemaïson, L. *Les ennemis animaux des plantes cultivées et des forêts*, 3 vols. Editions SEP, Paris, France (1962), 592 pp.
- 50 Sanchez, V.G.A. *Empoasca* spp. and *Macrosiphum euphorbiae* in sweet potato of Rimac and Canete Valleys (Peru). *Revista Peruana de Entomologia* **31** (1988) 117–118.
- 51 Melichar, L. *Cicadinen (Hemiptera – Homoptera) von Mitteleuropa*. Felix L. Dames, Berlin, Germany (1895) 364 pp.
- 52 Kaltenbach, J.H. *Die Pflanzenfeinde aus der Klasse der Insecten*. Thienemann, Stuttgart, Germany (1874) 848 pp.
- 53 Abdel-Fattah, M.I., Hendi, A. & El-Said, A. Abundance of *Bemisia tabaci* (Genn.) associated with common weeds in tomato fields at Shebin El-Kom region, Egypt (Homoptera: Aleyrodidae). *Bulletin de la Société Entomologique d'Égypte* **65** (1984) 109–117.
- 54 Legg, J.P. Host-associated strains within Ugandan populations of the whitefly *Bemisia tabaci* (Genn.), (Hom., Aleyrodidae). *Journal of Applied Entomology* **120** (1996) 523–527.
- 55 Börner, C. Europae centralis aphides, Part 1. In *Mitteilungen der Thüringischen Botanischen Gesellschaft* No. 4, Suppl. 3. Weimar, German Democratic Republic (1952) pp. 1–260.
- 56 Börner, C. Europae centralis aphides, Part 2. In *Mitteilungen Thüringischen Botanischen Gesellschaft*, No. 4, Suppl. 3. Weimar, German Democratic Republic (1952) pp. 261–484.
- 57 Borror, D.J., DeLong, D.M. & Triplehorn, C.A. *An introduction to the study of insects*, 4th ed. Holt, Rinehart & Winston, New York (1976) 852 pp.
- 58 Therrien, P. Individual food choices by foragers from the species *Acromyrmex octospinosus* (Reich), the leaf-cutting ant. *Memoirs of the Entomological Society of Canada* **146** (1988) 123–130.
- 59 Golberg, A.M., Avigdor-Avidor, H. & Nuriel, E. Insecticide control of a white grub, *Maladera matrida*, on sweet potato. *Phytoparasitica* **17** (1989) 175–184.
- 60 Balachowsky, A.S. Tome I. Coléoptères. In *Entomologie appliquée à l'agriculture*, Vol. 2. Masson, Paris, France (1963).
- 61 Rolston, L.H., Barlow, T., Jones, A. & Hernandez, T. Potential of host plant resistance in sweet potato (*Ipomoea batatas*) for control of a white grub, *Phyllophaga ephelida* (Coleoptera: Scarabaeidae). *Journal of the Kansas Entomological Society* **54** (1981) 378–380.
- 62 Hering, E.M. *Bestimmungstabellen der Blattminen von Europa einschließlich des Mittelmeerbeckens und der Kanarischen Inseln*. Vol. 1. Pflanzengattungen A–L. Erzeuger – Nr. 1-3133. W. Junk, The Hague, Netherlands (1957) pp. 1–648.
- 63 Medail, F., Ponel, P. & Barbero, M. Humid meadows of Garde and Pradet: their role in maintaining botanical and entomological diversity in the department of Var (France). *Bulletin de la Société Linnéenne de Provence* **45** (1994) 49–68.
- 64 Lohse, G.A. & Lucht, W.H. *Die Käfer Mitteleuropas, Suppl. 2*. Goecke & Evers, Krefeld, Germany (1992).
- 65 Reiter, E. *Fauna germanica. Die Käfer des Deutschen Reiches*, Vol. 5. K.G. Lutz, Stuttgart, Germany (1911).
- 66 Seal, D.R., McSorley, R. & Chalfant, R.B. Seasonal abundance and spatial distribution of wireworms (Coleoptera: Elateridae) in Georgia sweet potato fields. *Journal of Economic Entomology* **85** (1992) 1802–1808.
- 67 Jones, A., Schalk, J.M. & Dukes, P.D. Control of soil insect injury by resistance in sweet potato. *Journal of the American Society for Horticultural Science* **112** (1987) 195–197.
- 68 Seal, D.R. & Chalfant, R.B. Bionomics of *Conoderus rudis* (Coleoptera: Elateridae): Newly reported pest of sweet potato. *Journal of Economic Entomology* **87** (1994) 802–809.
- 69 Schalk, J.M., Jones, A., Dukes, P.D. & Burnham, K.P. Responses of soil insects to mixed and contiguous plantings of resistant and susceptible sweet potato cultivars. *HortScience* **27** (1992) 1089–1091.
- 70 Abdel-Rahman, H.A., Soliman, Z.A. & Ali, M.F. Ecological studies on the black carpet beetle, *Antageneus scalaris* Pic (Coleoptera: Dermestidae). *Bulletin de la Société Entomologique d'Égypte* **63** (1980–1981) 243–252.
- 71 Goncalves, P.A.S. Survey of insects associated with sweet potato, *Ipomoea batatas*, with water traps, in Ituporanga, Santa Catarina. *Anais da Sociedade Entomológica do Brasil* **26** (1997) 199–203.
- 72 Porta, A. *Fauna Coleopterorum Italica*, Vol. 5. Piacentino, Piacenza, Italy (1923–32).
- 73 Schuester, M. Blister beetle in Paraguay – a potential biological control agent. *Tropical Pest Management* **33** (1987) 241.
- 74 Fuller, C. The bindweed gall-marker (*Nupserha apicalis* F.). *Journal of the Department of Agriculture of the Union of South Africa* **8** (1914) 242–244.
- 75 Beeson, C.F.C. & Bhatia, B.M. On the biology of the Cerambycidae (Coleoptera). *Indian Forest Research* **5** (1939) 1–235.
- 76 Smith, J.B. Insects injurious to sweet potatoes in New Jersey. *New Jersey Agricultural Experimental Station Bulletin* **229** (1910) 16.
- 77 Wilcox, J.A. Leaf beetles of Ohio (Chrysomelidae: Coleoptera). *Ohio Biological Survey Bulletin* **43** (1954) 353–506.
- 78 Visalakshi, A., Santhakumari, K., Koshy, G. & Nair, M.R.G.K. Biological studies on *Aspidomorpha furcata* (Chrysomelidae: Cassidinae: Coleoptera). *Entomon* **5** (1980) 167–170.
- 79 Gressitt, J.L. The tortoise beetles of China (Chrysomelidae: Cassidinae). *Proceedings of the Californian Academy of Science* **27** (1952) 433–591.
- 80 Gressitt, J.L. & Kimoto, S. The Chrysomelidae (Coleoptera) of China and Korea. *Pacific Insects Monograph* **1B** (1963) 1020 pp.
- 81 Balsbaugh, E.U., Jr. & Riley, E.G. Two foreign tortoise beetles newly recorded from the United States (Coleoptera: Chrysomelidae: Cassidinae). *Coleopterists Bulletin* **34** (1980) 175–76.
- 82 Barwal, R.N. New records of insects of sweet potato, *Ipomoea batatas* Poiret in the northeastern region. *Journal of Insect Science* **5** (1992) 95.
- 83 Brown, W.J. Canadian species of *Calligrapha*. *Canadian Entomologist* **77** (1945) 117–133.

- 84 Freude, H., Harde, K.W. & Lohse, G.A. *Die Käfer Mitteleuropas*, Vol. 9. Goecke & Evers, Krefeld, German Federal Republic (1966) 299 pp.
- 85 Singh, J.P. & Sidhu, P.K. Life history of *Cassida indicola* Duvivier (Coleoptera: Chrysomelidae) – a pest on two species of *Convolvulus*. *Uttar Pradesh Journal of Zoology* **12** (1992) 40–44.
- 86 Virkki, N., Santiago Blay, J.A. & Riley, E.G. Chromosomes of Puerto Rican *Hispininae* and *Cassidinae* (Coleoptera: Chrysomelidae). *Coleopterists Bulletin* **46** (1992) 29–42.
- 87 Chittenden, F.H. The Argus tortoise beetle. *Journal of Agricultural Research* **27** (1924) 43–51.
- 88 Smith, R.C. A preliminary report on the insects attacking bindweed with special reference to Kansas. *Transactions of the Kansas Academy of Science* **41** (1938) 183–191.
- 89 Selleck, G.W. Biological control of perennial bindweeds with Argus tortoise beetle. In *Proceedings, 1978 British Crop Protection Conference – Weeds* (1978) pp. 499–502.
- 90 Balsbaugh, E.U., Jr., Frey, R.D., Scholl, C.G. & Anderson, A.W. Insects for weed control: status in North Dakota. *North Dakota Farm Research* **39** (1981) 3–7.
- 91 Menory-Ortega, J.G. Informe del entomólogo – patólogo. *Memorias de la Secretaria de Agricultura y Comercio de la República Dominicana* 1932, Santo Domingo (1934) pp. 117–133.
- 92 Reid, C.A.M. & Storey, R.I. Redescription of adult and larva of *Colasposoma sellatum* Baly (Coleoptera: Chrysomelidae: Eumolpinae): a pest of sweet potato in Australia. *Journal of Natural History* **27** (1993) 669–681.
- 93 Teng, H.J., Waddill, V., Slansky, F. & Strayer, J. Performance and host preference of adult banded cucumber beetles, *Diabrotica balteata*, when offered several crops. *Journal of Agricultural Entomology* **1** (1984) 330–338.
- 94 Schalk, J.M. & Creighton, C.S. Influence of sweet potato cultivars in combination with a biological control agent (Nematoda: *Heterorhabditis heliothidis*) on larval development of the banded cucumber beetle (Coleoptera: Chrysomelidae). *Environmental Entomology* **18** (1989) 897–899.
- 95 Schalk, J.M. Multiple insect resistance in sweet potato. In *Proceedings, Sweet Potato Weevil Workshop*, Southeastern Branch, Entomological Society of America, New Orleans, USA (1984) pp. 56–65.
- 96 Jolivet, P. Notes systématiques sur les chrysomélides marocains (coléoptères 2ième note). *Bulletin de la Société des Sciences Naturelles et Physiques du Maroc* **46** (1967) 305–394.
- 97 Rosenthal, S.S. & Carter, J. Host specificity and biology of *Galeruca rufa*, a potential biological control agent for field bindweed. *Environmental Entomology* **6** (1977) 155–158.
- 98 Rosenthal, S.S. & Hostettler, N. *Galeruca rufa* (Col.: Chrysomelidae) seasonal life history and effect of its defoliation on its host plant, *Convolvulus arvensis* (Convolvulaceae). *Entomophaga* **25** (1980) 383–390.
- 99 Kismali, S. & Madanlar, N. The role of Chrysomelidae (Coleoptera) species for the biological control of weeds and the status of the species in Izmir. In *Proceedings, 2nd Turkish National Congress of Biological Control* (1990) pp. 299–308.
- 100 Doguet, S. Coléoptères, Chrysomelidae, Alticinae. In *Faune de France 2*. Federation Francaise des Sociétés de Sciences Naturelles, Paris, France (1994) 694 pp.
- 101 Zorina, L.M. Some data on the biology of convolvulus flea-beetles. *Bulletin of the Institute of Zoology and Applied Phytopathology* **7** (1939) 34–43. [in Russian]
- 102 Jolivet, P. Les Chrysomeloidea (Coleoptera) des îles Baléares *Memoires de l'Institut Royal des Sciences Naturelles de Belgique, Ser. 2*. **50** (1953) 1–88.
- 103 Mohr, K.H. Bestimmungstabelle und Faunistik der mitteleuropäischen *Longitarsus* Arten. *Entomologische Blätter* **58** (1962) 55–118.
- 104 Mohr, K.H. 88. Familie: Chrysomelidae. In *Die Käfer Mitteleuropas*, Vol. 9, Freude, H.K., Harde, W. & Lohse, G.A. (Eds) Goecke & Evers, Krefeld, German Federal Republic (1966) 299 pp.
- 105 Shapiro, D.S. A review of the flea beetles (Coleoptera, Halticinae) of Dagestan and neighboring regions of the Caspian lowland. *Entomological Review* **48** (1969) 162–167.
- 106 Lopatin, I.K. & Tadzhibayev, M. The leaf beetles (Coleoptera, Chrysomelidae) of the low-mountain areas of Tajikistan. *Entomological Review* **51** (1972) 355–358.
- 107 Král, J. Ergebnisse der zoologischen Forschungen von Dr. Z. Kaszab in der Mongolei. 329. Alticinae III. (Coleoptera). *Folia Entomologica Hungarica, Supplement* **26** (1973) 111–133.
- 108 Lopatin, I.K. *Leaf Beetles (Chrysomelidae) of Central Asia and Kazakhstan*. Nauka, Leningrad, USSR (1977) 416 pp.
- 109 Furth, D.G. Zoogeography and host plants of *Longitarsus* in Israel, with description of six new species (Coleoptera: Chrysomelidae). *Israel Journal of Entomology* **13** (1979) 79–124.
- 110 Warchalowski, A. Chrzaszczki – Coleoptera, stonkowate – Chrysomelidae, podrodziny: Halticinae, Hispininae i Cassidinae. In *Klucze do oznaczania owadów Polski*, Part XIX, Issues 94b & 94c, Państwowe Wydawnictwo Naukowe, Warszawa, Poland (1978) 157 pp.
- 111 Shute, S.L. Wing polymorphism in British species of *Longitarsus* beetles (Chrysomelidae: Alticinae). *Systematic Entomology* **5** (1980) 437–448.
- 112 Lopatin, I.K. Chrysomelidae aus dem Himalaya (Coleoptera). *Entomologia Brasiliensia* **9** (1984) 328–329.
- 113 Heikertinger, F. & Ciski, E. Chrysomelidae: Halticinae. In *Coleopterorum catalogus*, Vol. 24, Parts 166 & 169, Schenkling, S. (Ed) W. Junk, The Hague, Netherlands (1940) 635 pp.
- 114 Liebmann, W. *Käferfunde aus Mitteleuropa einschliesslich des österreichischen*. Arnstadt, Alpen, German Democratic Republic (1955) 165 pp.
- 115 Nonveiller, G. *Stetni buvaci kulturnog i drugog korisnog bilja Srbije*. Institut za Zastitu Bilja, Beograd, Yugoslavia, special publication (1960) 56 pp. [in Serbian]
- 116 Regalin, R. New distributional data on some Italian species of *Alticinae* (Coleoptera, Chrysomelidae). *Bollettino della Società Entomologica Italiana* **110** (1978) 202–204.
- 117 Fogato, W. & Loenardi, C. Coleotteri crisomelidi della brughiera di Rovasenda (Piemonte). In *Quaderni sulla "Struttura delle zoocenosi terrestri" – 1. La brughiera pedemontana*, report (1980) pp. 25–73.
- 118 Maw, M.G. *Convolvulus arvensis* L., field bindweed (Convolvulaceae). In *Biological control programmes*

- against insects and weeds in Canada 1969–1980, Kelleher, J.S. & Hulme, M.A. (Eds) Commonwealth Agricultural Bureaux, London (1981) pp. 155–157.
- 119 Capinera, J.L. Consumption of sugar beet foliage by the palestriped flea beetle. *Journal of Economic Entomology* **71** (1978) 301–303.
- 120 Ware, G.W. & McCollum, J.P. *Producing vegetable crops*. Interstate Printers and Publishers Inc., USA (1968) 599 pp.
- 121 Parrella, M.P. & Kok, L.T. Bindweeds and their potential for biological control. *Virginia Journal of Science* **26** (1975) 44.
- 122 Teran, A.L. & Kingsolver, J.M. *Revision del genero Megacerus (Coleoptera: Bruchidae)*. Ministerio de Cultura y Education, Opera Lilloana, Tucuman, Argentina (1977) 287 pp.
- 123 Wang, R. & Kok, L.T. Host specificity of *Megacerus discoidus* (Coleoptera: Bruchidae) and its impact on hedge bindweed, *Calystegia sepium*. *Environmental Entomology* **15** (1986) 834–838.
- 124 Wang, R. *Megacerus discoidus, a potential biological control agent of hedge bindweed, Calystegia sepium in southwestern Virginia*. Virginia Polytechnic Institute and State University, Blacksburg, Virginia, USA (1985) 94 pp.
- 125 Schlising, R.A. Seed destruction of California morning glories (Convolvulaceae: *Calystegia*) by bruchid beetles. *Madrono* **27** (1980) 1–16.
- 126 Decelle, J.A. New species of *Spermophagus*: Coleoptera, Bruchidae, Amblyocerinae, of Turkey. *Bulletin et Annales de la Société Royale Belge d'Entomologie* **118** (1982) 31–34.
- 127 Decelle, J. Le genre *Spermophagus* Schönherr en Europe occidentale (Col.: Bruchidae: Amblyocerinae). *Bulletin de la Société Entomologique de France* **88** (1983) 235–241.
- 128 Wendt, H. Beiträge zur Insektenfauna der DDR: Coleoptera – Bruchidae (Chrysomeloidea). II. Bestimmungstabellen der heimischen Arten. *Mitteilungen aus dem Zoologischen Museum in Berlin* **64** (1988) 311–318.
- 129 Hoffmann, A. Coléoptères, bruchides et anthribides. In *Faune de France* 44, Lechevalier, Paris, France (1945) 184 pp.
- 130 Wendt, H. On the fauna of Bruchidae of Mongolia (Coleoptera, Chrysomeloidea). *Mitteilungen aus dem Zoologischen Museum in Berlin* **61** (1985) 279–285.
- 131 Borowiec, L. *Bruchidae – strąkowce (Insecta: Coleoptera)*. Państwowe Wydawnictwo Naukowe, Warszawa, Poland (1988) 225 pp.
- 132 Ishikawa, S.I., Toquenaga, Y. & Matsubayashi, T. Occurrence of *Spermophagus rufiventris* Boheman (Coleoptera, Bruchidae) in *Calystegia japonica* Choisy (Convolvulaceae) seeds. *Japanese Journal of Entomology* **62** (1994) 431–432.
- 133 Bagdasaryan, B.A. The seed beetles of the Armenia SSR and their relationships with plants, particularly Leguminosae. *Nauchnyje Trudy Erevanskovo Gosudarstvenovo Uniuersiteta* **16** (1941) 309–374.
- 134 Southgate, B.J. Biology of the Bruchidae. *Annual Review of Entomology* **24** (1979) 449–473.
- 135 Abdul-Rassoul, M.S., Othman, N.Y. & Dawah, H.A. Observation on the biology, host plants and distribution of Iraqi Bruchidae (Insecta, Coleoptera). *Journal of Biological Sciences and Research, Iraq* **17** (1986) 207–222.
- 136 Borowiec, L. The specific status of *Spermophagus variolosopunctatus* Gyllenhal, 1833 (Coleoptera, Bruchidae, Amblyocerinae), with a description of a new species. *Polskie Pismo Entomologiczne* **56** (1986) 161–164.
- 137 Baloch, G.M., Khan, A.G., Habib, R. & Zaffar, T. Laboratory testing and evaluation of insect enemies of halogeton and Russian thistle and research on biological control of weeds common to Pakistan and the United States. Sept. 1970–Sept. 1975. Pakistan Station, Commonwealth Institute of Biological Control, Rawalpindi, reports (1976) 66 pp.
- 138 Balachowsky, A.S. & Mesnil, L. *Les insectes nuisibles aux plantes cultivées*, Vol. 2. Etablissements Busson, Paris, France (1935–36).
- 139 Hoffmann, A. Coléoptères, curculionides, Part 1. In *Faune de France*, Lechevalier, Paris, France (1950) 486 pp.
- 140 Parker, B.L., Wolfe, G.W. & Abubaker, A. Occurrence of *Cylas formicarius* (F.) (Col., Apionidae) in central and southern Africa. *Journal of Applied Entomology* **114** (1992) 400–402.
- 141 Teli, V.S. & Salunkhe, G.N. Biology of sweet potato weevil. *Journal of Maharashtra Agricultural Universities* **19** (1994) 381–384.
- 142 Talekar, N.S. Characteristics of infestation of sweet potato by sweet potato weevil *Cylas formicarius* (Coleoptera: Apionidae). *International Journal of Pest Management* **41** (1995) 238–242.
- 143 Miyatake, T., Moriya, S., Kohama, T. & Shimoji, Y. Dispersal potential of male *Cylas formicarius* (Coleoptera: Brentidae) over land and water. *Environmental Entomology* **26** (1997) 272–276.
- 144 Hamilton, J.T. & Toffolon, R.B. Insect pests of sweet potato. *Agfacts* No. H8.AE.6 (1986) 3 pp.
- 145 Miyatake, T., Kawasaki, K., Kohama, T., Moriya, S. & Shimoji, Y. Dispersal of male sweetpotato weevils (Coleoptera: Curculionidae) in fields with or without sweet potato plants. *Environmental Entomology* **24** (1995) 1167–1174.
- 146 Anioke, S.C. Effect of time of planting and harvesting of sweet potato (*Ipomoea batatas* (L.) Lam.) on yield and insect damage in south-eastern Nigeria. *Entomon* **21** (1996) 137–141.
- 147 Simpson, S.E., Nigg, H.N., Coile, N.C. & Adair, R.A. *Diaprepes abbreviatus* (Coleoptera: Curculionidae): host plant associations. *Environmental Entomology* **25** (1996) 333–349.
- 148 Iwata, M., Shimabukuro, S., Makiguchi, S., Ishikawa, A. & Yasuda, K. Notes on adult emergence of West Indian sweet potato weevil, *Eusepes postfasciatus* (Fairmaire) (Coleoptera: Curculionidae) and emerged adult's physiological state. *Research Bulletin of the Plant Protection Service Japan* **30** (1994) 83–86.
- 149 Yasuda, K. Attractiveness of different plant organs of sweet potato and damaged tuber to West-Indian sweet potato weevil, *Eusepes postfasciatus* (Fairmaire) (Coleoptera: Curculionidae) in wind tunnel. *Japanese Journal of Applied Entomology and Zoology* **40** (1996) 160–161.
- 150 Yasuda, K. Occurrence of West Indian sweet potato weevil, *Eusepes postfasciatus* (Fairmaire) (Coleoptera: Curculionidae) and damage to sweet potato (*Ipomoea batatas* (L.) Lam.) fields. *Japanese Journal of Applied Entomology and Zoology* **41** (1997) 83–88.
- 151 Monteagudo, T.S., Machado, R.M., Grillo, R.H. & Suazo, G.M. A new host of *Pachnaeus litus* Germar (Coleoptera: Curculionidae). *Centro Agricola* **14** (1987) 57–60.

- 152 Spuler, A. *Die Schmetterlinge Europas*, Vol. 2. E. Schweizerbart, Stuttgart, Germany (1910) 523 pp.
- 153 Schutze, K.T. *Die Biologie der Kleinschmetterlinge unter besonderer Berücksichtigung ihrer Nahrungspflanzen und Erscheinungszeiten*. Frankfurt am Main, Germany (1931) 235 pp.
- 154 Hering, E.M. *Bestimmungstabellen der Blattminen von Europa einschließlich des Mittelmeerbeckens und der Kanarischen Inseln*. Vol. 2. *Pflanzengattungen M-Z. Erzeuger* – Nr. 3134-5551. W. Junk, The Hague, Netherlands (1957) pp. 649–1185.
- 155 Buszko, J. Studies on the mining Lepidoptera of Poland. X. Mining Lepidoptera of Torun and surrounding areas. *Acta Zoologica Cracoviensia* **33** (1990) 367–452.
- 156 Borkowski, A. Lepidoptera: Nepticulidae in the states of Berlin and Brandenburg, with comments on their treatment in the Red List of Endangered Animal Species. *Entomologische Nachrichten und Berichte* **38** (1994) 145–173.
- 157 Karsholt, O. & Razowski, J. (Eds) *The Lepidoptera of Europe (a distributional checklist)*. Apollo Books, Stenstrup, Denmark (1996) 380 pp.
- 158 Staudinger, O. Diagnosen nebst kurzen Beschreibungen neuer andalusischer Lepidopteren. *Entomologische Zeitung (Stettin)* **20** (1859) 211–259.
- 159 Lhomme, L. *Catalogue des Lépidoptères de France et de Belgique*, Vol. 2. Le Carriol, Douelle & Paris, France (1923–49)
- 160 Ahn, S.B. & Lim, S.E. Crop insect pests in Cheju Island: 1. Leaf feeding species on soybean and sweetpotato. *Research Reports of the Rural Development Administration (SUWEON)* **33** (1991) 46–50.
- 161 Marek, J., Lastůvka, A., Vavra, J., van der Wolf, H.W. Faunistic records from Czechoslovakia: Lepidoptera. *Acta Entomologica Bohemoslovaca* **89** (1992) 473–476.
- 162 Draghia, I. A study on a population of *Bedellia somnulentella* (Zeller) (Lepidoptera, Lyonetiidae) from Romania. *Travaux du Museum d'Histoire Naturelle "Grigore Antipa"* **15** (1974) 241–257.
- 163 Tawfik, M.F.S., Awadallah, K.T. & Shalaby, F.F. The life history of *Bedellia somnulentella* Zell. (Lepidoptera: Lyonetiidae). *Bulletin de la Société Entomologique d'Égypte* **60** (1976) 25–33.
- 164 Parrella, M.P. & Kok, L.T. The development and reproduction of *Bedellia somnulentella* on hedge bindweed and sweet potato. *Annals of the Entomological Society of America* **70** (1977) 925–928.
- 165 Heath, J. & Emmet, A.M. (Eds) Cossidae – Heliodinidae. In *The moths and butterflies of Great Britain and Ireland*, Vol. 2. Harley Books, London (1985) 460 pp.
- 166 Porter, C.E. Notas breves de entomologia agricola. *Anales de Zoologia Aplicada* **8** (1921) 20–21.
- 167 Liu, Y. Chinese *Ethmia* (Lepidoptera: Ethmiidae) in classification, distribution and numerical taxonomy. *Entomotaxonomia* **2** (1980) 267–284.
- 168 Bengtsson, B.A. Scythrididae. In *Microlepidoptera of Europe* 2, Heumer, P., Karsholt, O. & Lyneborg, L. (Eds) Apollo Books, Stenstrup, Denmark (1997) 301 pp.
- 169 Sanchez, V.G.A. & Redolfi de Yhuiza, I. Four microlepidopterous pest of sweet potato and their biological control in Rimac and Canete Valleys, (Peru). *Revista Peruana de Entomologia* **31** (1988) 113–116.
- 170 Gapasin, D.P. & Rebadulla, L.Z. Biology of the black leaf folder (*Brachmia* sp.) and green leaf folder (*Psara hipponalis*) of sweet potato. *Annals of Tropical Research* **3** (1981) 37–50.
- 171 Walsingham, T. de G. Microlepidoptera of Tenerife. Proceedings of the Zoological Society of London (1908) 1907(2), 911–1034.
- 172 Spuler, A. *Die Schmetterlinge Europas*, Vol. 1. E. Schweizerbart, Stuttgart, Germany (1908) 385 pp.
- 173 Setokuchi, O., Nakagawa, K. & Kobayashi, M. Food consumption of three major sweet potato defoliators, *Aedia funesta* Linne, *Agrius convolvuli* Linne, and *Brachmia triannulella* (Herrich-Schäffer). *Japanese Journal of Applied Entomology and Zoology* **30** (1986) 93–98.
- 174 Eckstein, K. Die Kleinschmetterlinge Deutschlands. In *Die Schmetterlinge Deutschlands*, Vol. 5. K.G. Lutz, Stuttgart, Germany (1933) 223 pp.
- 175 Laštůvka, Z. & Laštůvka, A. *An illustrated key to European Sesiidae*. Faculty of Agronomy MUA, Brno, Czech Republic (1995) 175 pp.
- 176 Bradley, J.D., Tremewan, W.G. & Smith, A. Cochyliidae and Tortricidae: Tortricinae. In *British tortricoid moths*. Ray Society, London (1973) 251 pp.
- 177 Balachowsky, A.S. Lepidoptères. In *Entomologie appliquée à l'agriculture*, Vol. 2. Masson, Paris, France (1966).
- 178 Hannemann, H.J. Kleinschmetterlinge oder Microlepidoptera. I. Die Wincler (Tortricidae). In *Die Tierwelt Deutschlands und der angrenzenden Meeresteile nach ihren Merkmalen und nach ihrer Lebensweise*, 41, Dahl, M. & Bischoff, H. (Eds) Gustav Fischer, Jena, German Democratic Republic (1961) 233 pp.
- 179 Razowski, J. Motyle – Lepidoptera, zwojtkowki – Tortricidae, wstep oraz podroziny Tortricinae i Sparganothinae. In *Klucze do oznaczania owadów Polski*, Part XXVII, Issue 41b, Państwowe Wydawnictwo Naukowe, Warszawa, Poland (1969) 131 pp.
- 180 de Graaf, H.W. Eine diagnostische beschrijving gemaakt van europesche Pterophoridae. *Tijdschrift voor Entomologie* **2** (1859) 35–57.
- 181 Amsel, H.G. Weitere Mitteilungen über palaestinensische Lepidopteren. *Veröffentlichungen Überseemuseum Bremen* **1** (1935) 225–277.
- 182 Gozmány, L.A. Microlepidoptera IV. In *Fauna Hungariae*, Akademia Kiado, Budapest, Hungary (1962) 298 pp.
- 183 Hannemann, H.J. Kleinschmetterlinge oder Microlepidoptera. III. Federermotten (Pterophoridae), Gespinstmotten (Yponomeutidae), Echte Motte (Tineidae). In *Die Tierwelt Deutschlands und der angrenzenden Meeresteile nach ihren Merkmalen und nach ihrer Lebensweise*, 63, Dahl, M. & Bischoff, H. (Eds) Gustav Fischer, Jena, German Democratic Republic (1977) 274 pp.
- 184 Parrella, M.P. & Kok, L.T. Bionomics of *Oidaematophorus monodactylus* on hedge bindweed in southwestern Virginia. *Annals of the Entomological Society of America* **71** (1978) 1–4.
- 185 Emmet, A.M. (Ed) A field guide to the smaller British Lepidoptera. British Entomological and Natural History Society, London (1979) 271 pp.
- 186 Parrella, M.P. & Kok, L.T. *Oidaematophorus monodactylus* as a biological control agent of hedge bindweed: Development of a rearing program and cost analysis. *Journal of Economic Entomology* **72** (1979) 590–592.

- 187 Buszko, J. A review of Polish Pterophoridae. *Polskie Pismo Entomologiczne* **56** (1986) 273–315.
- 188 Nel, J. On the first state of *Pterophorus (sensu stricto)* in France: fifth contribution to the knowledge of the biology of Pterophoridae from southern France (Lepidoptera, Pterophoridae). *Alexanor, Supplement* **15** (1987) 29–32.
- 189 Arenberger, E. Pterophoridae from Kashmir (India). *Zeitschrift der Arbeitsgemeinschaft Oesterreichischer Entomologen* **40** (1988) 23–32.
- 190 Arenberger, E. & Jaksic, P. Pterophoridae (Insecta, Lepidoptera). In *Fauna Durmitora*, Part 4. Montenegrin Academy of Sciences and Arts, Titograd, Yugoslavia (1991) pp. 225–242.
- 191 Gielis, C. Pterophoridae. In *Microlepidoptera of Europe*, Vol. 1. Huemer, P., Karsholt, O. & Lyneborg, L. (Eds) Apollo Books, Stenstrup, Denmark (1996) 220 pp.
- 192 Yano, K. Taxonomic and biological studies of Pterophoridae of Japan (Lepidoptera). *Pacific Insects* **5** (1963) 65–209.
- 193 Meyrick, E. *A revised handbook of British Lepidoptera*. Watkins & Doncaster, London (1927) 914 pp.
- 194 Hrubý, K. *Prodromus lepidopter slovenska – Prodromus lepidopterarum slovaciae*. SAV, Bratislava, Czechoslovakia (1964) 962 pp.
- 195 Buzsko, J. Motyle – Lepidoptera, przeglądki – Thyrididae & piórolotky – Pterophoridae. In *Klucze do oznaczania owadów Polski*, Part XXVII, Issues 43 & 44, Państwowe Wydawnictwo Naukowe, Warszawa (1979) 139 pp.
- 196 Ghani, M.A., Baloch, G.M., Khan, A.G., Habib, R. & Zaffar, T. Laboratory testing and evaluation of insect enemies of halogeton and Russian thistle and research on biological control of weeds common to Pakistan and the United States. Pakistan Station, Commonwealth Institute of Biological Control, Rawalpindi, final report (1975) 59 pp.
- 197 Hampson, G.F. *Moths*, Vol 4. In *The fauna of British India – including Ceylon and Burma*. Taylor & Francis, London (1896) 594 pp.
- 198 Camprag, D. & Sekulic, R. Harmful fauna on sunflowers in Yugoslavia and special review of the pests in Vojvodina Province. *Savremena Poljoprivreda* **35** (1987) 79–91.
- 199 Sannino, L., Espinosa, B. & Balbiani, A. Comparative morphological study on the immature stages of the Lepidoptera tobacco pests. 2. Larvae. *Bollettino del Laboratorio di Entomologia Agraria "Filippo Silvestri"* **50** (1993) 23–91.
- 200 Vergara de Sanchez, C.E. & Sanchez, G.A. Biological cycle of *Microthyris anormalis* (Gn.) (Lepidoptera, Pyralidae). *Revista Peruana de Entomologia* **31** (1988) 107–109.
- 201 Talekar, N.S. & Cheng, K.W. Nature of damage and sources of resistance to sweetpotato vine borer (Lepidoptera: Pyralidae) in sweet potato. *Journal of Economic Entomology* **80** (1987) 788–791.
- 202 Talekar, N.S., Hu, W.J., Su, F.C., Yeh, W.B. & Lyou, Y.H. Mating and sex pheromone related studies with sweetpotato vine borer (Lepidoptera: Pyralidae). *Chinese Journal of Entomology* **12** (1992) 277–284.
- 203 Goater, B. *British pyralid moths*. Harley Books, London (1986) 175 pp.
- 204 Pinhey, E. *Hawk moths of central and southern Africa*. Longmans, Green & Co., London (1962) 139 pp.
- 205 Newman, L.H. *Hawk moths of Great Britain and Europe*. Cassell, London (1965) 148 pp.
- 206 Allan, P.B.M. *Larval foodplants*. Watkins & Doncaster, London (1949) 126 pp.
- 207 Nair, M.R.G.K. *Insects and mites of crops in India*. ICAR, New Delhi, India (1975) 404 pp.
- 208 Nayar, K.K., Ananthkrishnan, T.N. & David, B.V. *General and applied entomology*. Tata McGraw-Hill, New Delhi, India (1976) 589 pp.
- 209 Heath, J. & Emmet, A.M. (Eds) Sphingidae – Noctuidae, Noctuidae and Hadeninae. In *The moths and butterflies of Great Britain and Ireland*, Vol. 9. Harley Books, London (1979) 288 pp.
- 210 Pittaway, A.R. The 'Arabian sphinx'. *Ahlan Wasahlan* **11** (1987) 42–45.
- 211 Koch, M. *Wir bestimmen Schmetterlinge*. Neumann-Neudamm, Leipzig, German Democratic Republic (1988) 792 pp.
- 212 Nilsson, L.A. & Rabakonandrianina, E. Hawk moth scale analysis and pollination specialization in the epilithic Malagasy endemic *Aerangis ellisi* (Reichenb. fil.) Schltr. (Orchidaceae). *Botanical Journal of the Linnean Society (London)* **97** (1988) 49–61.
- 213 Govindan, R., Sarayanaswamy, T.K., Gururajarao, M.R. & Satenahalli, S.B. Insect infesting wild mung (*Vigna vexillata*) in India. *Environment and Ecology* **7** (1989) 513.
- 214 Shaw, S.S., Verma, R.S., Badaya, A.K., Mandloi, K.C. & Bhalla, P.L. An outbreak of hawk moth in Nimar Valley of Madhya Pradesh. *Indian Journal of Pulses Research* **2** (1989) 93.
- 215 Whitcombe, R.P. & Erzinclioğlu, Y.Z. The convolvulus hawk moth, *Agrius convolvuli* (Lepidoptera: Sphingidae) and its parasite *Zygobothria ciliata* (Diptera: Tachinidae) in Oman. *Journal of Oman Studies* **10** (1989) 77–84.
- 216 Wiedemann, H.J. & Köhler, J. *Nachtfalter: Spinner und Schwärmer*. Naturbuch, Augsburg, Germany (1996) 205 pp.
- 217 Bell, T.R.D. & Scott, I.A. *Moths*, Vol. 5, *Sphingidae*. In *The fauna of British India – including Ceylon and Burma*. Taylor & Francis, London (1937) 538 pp.
- 218 Wysoki, M. & Izhar, Y. Biological data on *Apanteles cerialis* (Hymenoptera: Braconidae), a parasite of *Boarmia selenaria* (Lepidoptera: Geometridae). *Phytoparasitica* **9** (1981) 19–26.
- 219 Ahn, S.B., Im, D.J., Kim, I.S. & Cho, W.S. Foliage-feeding lepidopterous pests on apple trees in Suwon (South Korea). *Research Reports of the Rural Development Administration (SUWEON)* **31** (1989) 27–33.
- 220 Kulkarni, N., Joshi, K.C. & Sambath, S.A. New report of *Ascotis selenaria imparata* Walk. (Lepidoptera: Geometridae) as a pest of *Pongamia pinnata* (L.) Pierre. *Indian Forester* **121** (1995) 239–240.
- 221 Kulkarni, N., Kalia, S., Sambath, S. & Joshi, K.C. First report of *Ascotis selenaria imparata* Walk. (Lepidoptera: Geometridae) as a pest of *Moringa pterigosperma* Gertn. *Indian Forester* **122** (1996) 1075–1076.
- 222 Ando, T., Ohtani, K., Yamamoto, M., Miyamoto, T. & Qin, X. Sex pheromone of Japanese giant looper, *Ascotis selenaria cretacea*: identification and field tests. *Journal of Chemical Ecology* **23** (1997) 2413–2423.
- 223 Seppänen, E.J. *The food-plants of the larvae of the Macrolepidoptera of Finland*. Werner Söderström Osakeyhtiö, Porvoo & Helsinki, Finland (1970) 179 pp.

- 224 Heath, J. & Emmet, A.M. (Eds) Noctuidae (Cuculliinae to Hypeninae) and Agaristidae. In *The moths and butterflies of Great Britain and Ireland*, Vol. 10. Harley Books, London (1983) 459 pp.
- 225 Blasche, P. *Raupenkalender für das mitteleuropäische Faunengebiet. Nach den Futterpflanzen geordnet*. Alfred Kernen, Stuttgart, German Federal Republic (1955) 149 pp.
- 226 Buzsko, J. Motyle – Lepidoptera, sówki – Noctuidae. In *Klucze do oznaczania owadów Polski*, Part XXVII, Issue 53e, Państwowe Wydawnictwo Naukowe, Warszawa, Poland (1983) 170 pp.
- 227 Seitz, A. (Ed) *The Macrolepidoptera of the world*, Section I, *The Macrolepidoptera of the Palaearctic region*, Vol. 3, *The noctuid moths*. Alfred Kernen, Stuttgart, Germany (1914) 511 pp.
- 228 Maceljski, M. & Balarin, I. On knowledge of polyphagy and its importance for the silver-Y moth (*Autographa gamma* L.). *Acta Entomologica Jugoslavica* **8** (1972) 39–54.
- 229 Napiorkowska, J., Kowalik, J., Machowicz, Z. & Stefaniak, Z. Evaluation of parasitic insects and entomophagous fungi as natural controls of noctuid larvae (Lepidoptera, Noctuidae) occurring on plantations of cabbages and sugar beets. *Polskie Pismo Entomologiczne* **56** (1986) 687–700.
- 230 Forster, W. & Wohlfahrt, T.A. Eulen (Noctuidae). In *Die Schmetterlinge Mitteleuropas*, Vol. IV. Franckh, Stuttgart, German Federal Republic (1971) 329 pp.
- 231 Cabello, T. Natural enemies of noctuid pests (Lepidoptera: Noctuidae) on alfalfa, corn, cotton and soybean crops in southern Spain. *Journal of Applied Entomology* **108** (1989) 80–88.
- 232 Etman, A.A.M. On some factors influencing the population dynamics of *Spodoptera littoralis* (Boisd.), *Spodoptera exigua* (Hbn.), *Syngrapha circumflexa* (L.), *Autographa gamma* L., and *Heliothis armigera* (Hbn.) (Lepidoptera, Noctuidae) in Egypt. *Journal of Applied Entomology* **108** (1989) 182–190.
- 233 Singh, J., Sandhu, S.S., Singla, M.L. & Singh, J. Ecology of *Heliothis armigera* (Hub.) on chickpea in Punjab. *Journal of Insect Science* **3** (1990) 47–52.
- 234 Shi, Q., Liu, W., Shao, Z. & Jia, H. The distribution of *Helicoverpa armigera* on different crops in north China. *Sinozoologia, Supplement* **12** (1995) 36–39.
- 235 Kurir, A. Tests on foodplants of *Mamestra oleracea* (Lepidoptera Noctuidae), valid for Europe. *Zeitschrift für Angewandte Entomologie* **94** (1982) 93–98.
- 236 Blackford, M. & Dinan, L. The effects of ingested ecdysteroid agonists (20-hydroxyecdysone, RH5849 and RH5992) and an ecdysteroid antagonist (cucurbitacin B) on larval development of two polyphagous lepidopterans (*Acherontia atropos*) and (*Lacanobia oleracea*). *Entomologia Experimentalis et Applicata* **83** (1997) 263–76.
- 237 Tóth, P., Vráblová, M. & Cagaň, L. Importance of noctuid species for biological control of field bindweed (*Convolvulus arvensis* L.) and hedge bindweed (*Calystegia sepium* L.). In *Proceedings, 14th Slovak and Czech Plant Protection Conference*, SPU, Nitra, Slovakia (1997) pp. 301–302.
- 238 Rahn, R. Lepidoptera that prey on cabbage (*Brassica oleracea* var. *capitata*) plantings in western France. *Sciences Agronomiques Rennes* **2** (1983) 48–60.
- 239 Nikolov, N.K. Threshold of economic harmfulness of *Mamestra brassicae*, *Mamestra oleracea* and *Plusia gamma* (Lepidoptera: Noctuidae) in pepper. *Rastenievudni Nauki* **27** (1990) 94–99.
- 240 Ippolito, R. & Parenzan, P. *Hoplodrina ambigua* D. and S. and other Lepidoptera, Noctuidae in artichoke fields in Apulia (Italy). *Entomologica* **20** (1985) 147–158.
- 241 Valverde, C.A. & Sarmiento, M.J. Four host-plants influencing susceptibility of *Spodoptera eridania* to three insecticides. *Revista Peruana de Entomologia* **29** (1986) 61–64.
- 242 Mattana, A.L. & Foerster, L.A. Consumption and utilization of leaves of sweet potato and 'bracatinga' by larvae of *Spodoptera eridania* (Cramer, 1782) (Lepidoptera: Noctuidae). *Anais da Sociedade Entomologica do Brasil, Supplement* **17** (1988) 95–106.
- 243 Seppänen, E.J. *Suomen suurperhostoukkien ravintokasvit*. Werner Söderström Osakeyhtiö, Porvoo & Helsinki, Finland (1954) 416 pp.
- 244 Rosenthal, S.S. Host specificity of *Tyta luctuosa* (Lepidoptera: Noctuidae), an insect associated with *Convolvulus arvensis* (Convolvulaceae). *Entomophaga* **23** (1978) 367–370.
- 245 Felt, E.P. *Plant galls and gall-makers*. Comstock, Ithaca, New York (1940) 364 pp.
- 246 Nijveldt, W. *Gall midges of economic importance*, Vol. VIII. Crosby Lockwood, London (1950) 231 pp.
- 247 Spencer, K.A. *Agromyzidae (Diptera) of economic importance*. W. Junk, The Hague, Netherlands (1973) 418 pp.
- 248 Spencer, K.A. The Agromyzidae (Diptera) of Fennoscandia and Denmark. In *Fauna Entomologica Scandinavica*, Vol. 5. Scandinavian Science Press, Klampenborg, Denmark (1976) 606 pp.
- 249 Tóth, P., Vráblová, M. & Cagaň, L. First record of *Melanagromyza albocilia* (Agromyzidae, Diptera) from Slovakia. *Biologia (Bratislava)* **53** (1998) 604.
- 250 Spencer, K.A. A new agromyzid (Diptera) from Pakistan feeding as a stem-borer in *Convolvulus arvensis*. *Bulletin of Entomological Research* **61** (1971) 369–371.
- 251 Austin, D.F. The taxonomy, evolution, and genetic diversity of sweet potatoes and related wild species. In *Exploration and Maintenance and Utilization of Sweet Potato Genetic Resources, First Planning Conference*, Lima, International Potato Center (CIP), Peru (1988) pp. 27–59.
- 252 McClay, A.S., Littlefield, J.L. & Kashefi, J. Establishment of *Aceria malherbae* (Acari: Eriophyidae) as a biological control agent for field bindweed (Convolvulaceae) in the Northern Great Plains. *Canadian Entomologist* **131** (1999) 541–547.
- 253 Tóth, P., Cristofaro, M. & Cagaň, L. Seasonal biology of *Melanagromyza albocilia* Hendel (Diptera: Agromyzidae) and seasonal patterns of field bindweed infestation, under field conditions in Slovakia. *Entomologica Fennica* (2005) (in press).
- 254 Défago, G., Ammon, H.U., Cagaň, L., Draeger, B., Graeves, M.P., Guntli, D., Hoecke, D., Klimeš, L., Lawrie, J., Moënné-Loccoz, Y., Nicolet, B., Pflirter, H.A., Tabacchi, R. & Tóth, P. Towards the biocontrol of bindweeds with a mycoherbicide. *BioControl* **46** (2001) 157–173.
- 255 Spencer, K.A. The species–host relationship in the Agromyzidae (Diptera) as an aid to taxonomy. In *Proceedings, 12th International Congress of Entomology*, London (1965) pp. 101–102.
- 256 Hendel, F. 59. Agromyzidae. In *Die Fliegen der Palaearktischen Region*, Vol VI, Lindner, E. (Ed), E. Schweizerbart, Stuttgart, Germany (1938) 570 pp.

- 257 Soós, A. & Papp, L. (Eds) *Catalogue of Palaearctic Diptera. Micropezidae – Agromyzidae*, Vol. 9. Elsevier, Budapest, Hungary (1984) 460 pp.
- 258 Spencer, K.A. Some Agromyzidae (Diptera) from Israel. *Israel Journal of Entomology* **9** (1974) 141–157.
- 259 Vála, M. & Černý, M. Agromyzidae. In *Checklist of Diptera (Insecta) of the Czech and Slovak Republics*, Chvála, M. (Ed) Karolinum, Charles University Press, Prague, Czech Republic (1997) pp. 75–78.
- 260 LeSage, L. Notes on European *Longitarsus* species introduced in North America (Coleoptera: Chrysomelidae: Alticinae). *Canadian Entomologist* **120** (1988) 1133–1145.
- 261 Bridwell, J.C. Notes on the Bruchidae and their parasites in the Hawaiian Islands. In *Proceeding of the Hawaii Entomological Society* **3** (1918) 465–505.
- 262 Viggiani, G. Studies on the hymenopteran Chalcidoidea: 62. *Uscana spermophagi*, new species (Trichogrammatidae), parasite of *Spermophagus sericeus* (Coleoptera, Bruchidae). *Bollettino del Laboratorio di Entomologia Agraria “Filippo Silvestri”* **36** (1979) 51–54.
- 263 Rosenthal, S.S. European organisms of interest for the biological control of *Convolvulus arvensis* in the United States. In *Proceedings, 5th International Symposium on Biological Control of Weeds*, Brisbane, Australia, 22–27 July 1980 (1981) pp. 537–544.
- 264 Selleck, G.W. Biological control of perennial bindweeds (*Convolvulus* sp.) with Argus tortoise beetle in horticultural crops. In *Abstracts, 1979 meeting of the Weed Science Society of America* (1979) p. 76.
- 265 Tóth, P., Tóthová, M. & Cagáň, L. *Emmelina monodactyla* (Linnaeus, 1758) (Lepidoptera: Pterophoridae), its parasitization and potential as a biological control agent of field or lesser bindweed (*Convolvulus arvensis* L.). *Entomologist's Gazette* **54** (2003) 233–241.
- 266 Steinbauer, C.E. & Kushman, L.J. Sweet potato culture and diseases. *USDA Agriculture Handbook* **388** (1971) 1–74.
- 267 Bogdanov-Kačkov, N.N. & Tropkina, M.F. Pests of sweet potato and their quarantine importance. In *Pests and diseases of sweet potato*. State Association for Pest and Disease Control in Agriculture and Forestry in the USSR, Quarantine Administration, Moscow, Leningrad, USSR (1933) 5–217.
- 268 Parrella, M.P. & Kok, L.T. *Oidaematophorus monodactylus*: Oviposition and development on sweet potato and inundative releases on hedge bindweed. *Environmental Entomology* **7** (1978) 803–806.
- 269 Shorey, H.H. & Anderson, L.D. Biology and control of the morning glory leaf-miner, *Bedelia somnulentella* on sweet potatoes. *Journal of Economic Entomology* **53** (1960) 1119–1122.
- 270 Hinkley, A.D. Lepidopterous leaf-miners on sweet potato in Fuji. *Bulletin of Entomological Research* **53** (1963) 665–670.