

Biologically active peptides derived from egg proteins

Y. MINE* and J. KOVACS-NOLAN

Department of Food Science, University of Guelph, Guelph, Ontario N1G 2W1, Canada

*ymine@uoguelph.ca

Keywords: egg proteins; bioactive peptides; nutraceuticals; chronic disease; human health

Summary

Bioactive peptides are specific protein fragments that positively impact body's function or condition and ultimately may influence health. These peptides are inactive within the sequence of the parent protein and can be released during proteolysis or fermentation. These peptides may exert a number of different activities *in vivo*, affecting cardiovascular, endocrine, immune and nervous system in addition to nutrient utilization. Eggs have traditionally been recognized as an excellent source of protein, vitamins and minerals. Research in the past decade, however, has produced a substantial amount of evidence indicating that egg peptides may exert several diverse biological effects, above and beyond fulfilling basic nutritional requirements. Several biological activities have now been associated with egg proteins, including novel antimicrobial activities, immunomodulatory, anti-cancer, and anti-hypertensive activities, highlighting the importance of egg proteins in human health, and disease prevention and treatment. Continued research to identify new and existing biological functions of hen egg proteins will help to define new methods to further improve the value of eggs, as a source of numerous biologically active compounds with specific benefits for human and animal health, and secure their role in the therapy and prevention of chronic and infectious disease.

Introduction

Eggs consist of approximately 9.5% eggshell (including shell membrane), 63% albumen, and 27.5% yolk (Cotterill and Geiger, 1977). The main components are water (75%), proteins (12%), lipids (12%), as well as carbohydrates and minerals (Burley and Vadehra, 1989). The proteins are distributed throughout the egg, with the majority found in the egg yolk and egg white, and a small proportion in the eggshell and shell membrane (Sugino *et al.*, 1997). The lipids are found almost exclusively in the egg yolk, mainly in the form of lipoproteins (Burley and Vadehra, 1989). Several minerals have also been found in eggs, most of them in the eggshell. Carbohydrates are a minor egg component, present throughout the egg, both as free and conjugated forms, attached to proteins and lipids (Sugino *et al.*, 1997). Many diverse biological functions of egg proteins/peptides will be discussed here.

Egg white

The egg white, or albumen, makes up about 60% of the total egg weight of which water and protein are the major constituents (Sugino *et al.*, 1997). The egg white proteins include ovalbumin, which is the major protein, followed by ovotransferrin and ovomucoid. Other egg white proteins include ovomucin, which is responsible for the viscosity of the albumen, lysozyme, avidin, cystatin, ovomucoid, and ovomacroglobulin (ovostatin) (Sugino *et al.*, 1997).

ANTIMICROBIAL ACTIVITY

Egg white contains a number of proteins with demonstrated antimicrobial activities, which act as part of the natural defence system of the egg. Lysozyme, which exerts bacteriolytic activity by hydrolyzing the $\beta(1-4)$ linkage between N-acetylmuraminic acid and N-acetylglucosamine of peptidoglycan, the structural component of bacterial cell walls, has been studied at length, and has been applied as a natural food preservative (Losso *et al.*, 2000). It is most effective against Gram positive bacteria such as *Bacillus stearothermophilus*, *Clostridium tyrobutyricum*, and *Clostridium thermosaccharolyticum*, but this spectrum of activity can be broadened to include other spoilage and pathogenic organisms, as well as Gram negative bacteria, when used in conjunction with compounds such as EDTA, organic acids, or nisin (Losso *et al.*, 2000), or by coupling it with a hydrophobic carrier or compounds lethal to

the bacterial membrane (Ibrahim *et al.*, 2001). Studies have suggested that lysozyme may possess further antibacterial activity independent of its catalytic functions (Ibrahim *et al.*, 2001). Enzymatic hydrolysis of lysozyme has been found to enhance its activity, by exposing antibacterial portions of the protein (Pellegrini *et al.*, 1997; Ibrahim *et al.*, 2001), and producing peptides with antibacterial activity. Peptides corresponding to amino acid residues 98-112 (Pellegrini *et al.*, 2000), 98-108, and 15-21 (Mine *et al.*, 2004) possessed antimicrobial activity against *Escherichia coli* and *Staphylococcus aureus*. Lysozyme has also demonstrated antiviral activity. Oral and topical applications of lysozyme were found to be effective in preventing and controlling several viral skin infections, including herpes simplex and chicken pox (Sava, 1996), and Lee-Huang *et al.* (1999) found that chicken lysozyme also possessed activity against HIV type 1. Peptides produced by the enzymatic digestion of ovalbumin, one of the major proteins present in egg white, and their synthetic counterparts, were found to be strongly active against *Bacillus subtilis* and to a lesser extent against *Escherichia coli*, *Bordetella bronchiseptica*, *Pseudomonas aeruginosa*, and *Serratia marcescens*, as well as *Candida albicans* (Pellegrini *et al.*, 2004). Ovotransferrin, a member of the transferrin family, a group of iron-binding proteins widely distributed in various biological fluids, has the capacity to reversibly bind iron (Ibrahim, 2000). Its suggested function is as an iron scavenger, to prevent its use by microorganisms, and as an iron delivery agent (Abdallah *et al.*, 1999), and it has demonstrated antibacterial activity against a wide spectrum of bacteria, including *Pseudomonas* spp., *Escherichia coli*, *Streptococcus mutans* (Valenti *et al.*, 1983), *Staphylococcus aureus*, *Bacillus cereus* (Abdallah *et al.*, 1999) and *Salmonella enteritidis* (Baron *et al.*, 2000). Ovotransferrin has also been found to exert antibacterial activity by permeating bacterial outer membranes, reaching the inner membrane and causing the selective permeation of ions and dissipation of electrical potential (Aguilera *et al.*, 2003). A 92-amino acid ovotransferrin peptide, OTAP-92, was found to be capable killing Gram negative bacteria by crossing the bacterial outer membrane by self-promoted uptake, damaging the cytoplasmic membrane (Ibrahim *et al.*, 2000). It has also shown antiviral activity, against Marek's disease virus in chicken embryo fibroblasts (Giansanti *et al.*, 2002). Avidin, which possesses the unique ability to specifically bind the water soluble vitamin biotin, has been found to inhibit the growth of biotin-requiring bacteria and yeasts (Banks *et al.*, 1986). Avidin antimicrobial activity has also been attributed to its ability to bind to various Gram negative and Gram positive bacteria, including *Escherichia coli* K-12, *Klebsiella pneumoniae*, *Serratia marcescens*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and *Staphylococcus epidermidis* (Korpela *et al.*, 1984). Ovomucin-derived peptides, have demonstrated antiviral activity against Newcastle disease virus, bovine rotavirus, and human influenza virus *in vitro* (Tsuge *et al.*, 1996a,b, 1997a,b; Watanabe *et al.*, 1998a).

ANTI ADHESIVE PROPERTIES

The adhesion of microorganisms to host tissues is the first step in the infection process, in many cases mediated by an interaction between components on the surface of the microorganism and carbohydrates on the mucosal surface of the host (Sharon and Ofek, 2002). It has been suggested, then, that oligosaccharides and glycoconjugates, such as analogues of carbohydrates on the mucosal surface would competitively inhibit microorganism-carbohydrate adhesion on intestinal cells, thereby preventing microbial infection (Kobayashi *et al.*, 2004). Kobayashi *et al.* (2004) found, however, that enzymatic digestion of the egg white protein ovomucin, which is highly glycosylated, produced ovomucin glycopeptides (OGP), which possessed *Escherichia coli* O157:H7-specific binding sites, consisting of sialic acid. Based on these findings, it was suggested that OGP may be protective against *E. coli* O157:H7 infection *in vivo*, as well as a potentially novel probe for *E. coli* O157:H7 detection.

IMMUNOMODULATING ACTIVITY

The immune system responds to antigenic stimulation with a complex array of molecular events, involving antigen-presenting cells, B-cells, T-cells, and phagocytes. Cytokines play a significant role in regulating such immune responses (Wahn, 2003). Concomitant is their involvement in the pathology of a wide range of diseases and their potential in replacement and immunomodulatory therapy. Several egg white proteins and peptides have demonstrated immunomodulating activity. Ovalbumin has been found to induce the release of tumour necrosis factor (TNF) alpha in a dose-dependant manner *in vitro*, when modified with dicarbonyl methylglyoxyl (Fan *et al.*, 2003), and immunogenic ovalbumin peptides have been used to enhance immune responses for cancer immunotherapy (Vidovic *et al.*, 2002; Goldberg *et al.*, 2003; He *et al.*, 2003). Tezuka and Yoshikawa (1995) found that the phagocytic activity of macrophages was increased by the addition of ovalbumin peptides, OA 77-

84 and OA 126-134, derived from peptic and chymotryptic digestions of ovalbumin, respectively. Likewise, ovomucin peptides have shown macrophage-stimulating activity *in vitro* (Tanizaki *et al.*, 1997). Synthetic ovomucoid peptides have also demonstrated immunomodulating activity, inducing T-cell secretion of cytokines interleukin- (IL) 4, IL-10, IL-13, interferon- (IFN) gamma, and IL-6 (Holen *et al.*, 2001). Lysozyme has been shown to act as an immune-modulating and immune-stimulating agent. It has been shown, when combined with immunotherapy, to be effective in improving chronic sinusitis (Asakura *et al.*, 1990), and to normalize humoral and cellular responses in patients with chronic bronchitis (Sava, 1996). It was also found to enhance antibody production in hybridomas and lymphocytes, being termed an immunoglobulin production stimulating factor (Sugahara *et al.*, 2000), and to regulate and restore the immune responses in immune-depressed patients undergoing anti-cancer treatments (Sava, 1996). Furthermore, it has been suggested that the antibacterial activity of lysozyme might also occur via stimulation of macrophage phagocytic function, and that the hydrolysis products of peptidoglycan may act as an adjuvant or immunomodulator (Li-Chan and Nakai, 1989). Ovotransferrin is an acute phase protein in chickens, serum levels of which increase in inflammation and infections (Xie *et al.*, 2002). Xie *et al.* (2002) demonstrated that ovotransferrin can act as an immunomodulator, modulating macrophage and heterophil functions *in vitro*. Further immunomodulating effects of ovotransferrin have also been shown, including the inhibition of proliferation of mouse spleen lymphocytes (Otani and Odashima, 1997) and the enhanced phagocytic response of peripheral blood mononuclear cells and polymorphonuclear cells in dogs (Hirota *et al.*, 1995). Finally, research has suggested that cystatins may also be involved in inflammation and immune responses through the cytokine network, through mechanisms unrelated to the known protease inhibitory regions of the molecule (Kato *et al.*, 2000). Verdot *et al.* (1996, 1999) found that cystatin induced the synthesis of TNF- and IL-10, resulting in an upregulation of nitric oxide *in vitro* using mouse peritoneal macrophages. It was also found to upregulate the production of IL-6 by human gingival fibroblast cells and murine splenocytes, and the IL-8 production of gingival fibroblasts (Kato *et al.*, 2000).

ANTI-CANCER ACTIVITY

Lysozyme has been studied extensively as an anticancer agent, and has been shown to inhibit tumour formation and growth, both *in vitro* and *in vivo* when administered orally, in a number of experimental tumours, including lung carcinoma (Sava *et al.*, 1991; Das *et al.*, 1992; Pacor *et al.*, 1999). It was also found to enhance the efficacy of chemotherapy treatments (Sava *et al.*, 1995), and have a preventative effect when administered to normal mice (Das *et al.*, 1992). Evidence suggests that the anticancer effects of orally administered lysozyme may rely heavily upon the host-mediated immune response, including activation of the spleen and macrophages (Sava *et al.*, 1991), however, more recent data indicates that lysozyme may also exert action on the tumour cells themselves (Pacor *et al.*, 1999). Pronase-prepared glycopeptides of ovomucin have also demonstrated anti-tumour effects in a double grafted tumour system in mice, suggested to be related to the anti-angiogenic activity of ovomucin, inhibiting tumour growth (Oguro *et al.*, 2001).

ANTI-HYPERTENSIVE ACTIVITY

A vasorelaxing peptide, ovokinin (OA 358-365), was isolated by the peptic digestion of ovalbumin (Fujita *et al.*, 1995a). Ovokinin (2-7), a peptide produced by chymotrypsin digestion, and corresponding to OA 359-364, was also found to possess vasorelaxing activity (Matoba *et al.*, 1999). Both peptides were found to significantly lower the systolic blood pressure in spontaneously hypertensive rats, in a dose-dependent manner, when administered orally (Fujita *et al.*, 1995b). The replacement of amino acids in the ovokinin (2-7) peptide has resulted in enhanced anti-hypertensive activity, with the most potent derivative resulting in 100-fold more potent anti-hypertensive activity (Matoba *et al.*, 2001; Yamada *et al.*, 2002). Two angiotensin I converting enzyme (ACE)-inhibitory peptides were also identified in ovalbumin by peptic (OA 183-184) and tryptic (OA 200-218) digestions (Yoshikawa and Fujita, 1994). Miguel *et al.* (2004) examined peptides with angiotensin-converting enzyme (ACE)-inhibitory properties produced by enzymatic hydrolysis of crude egg white, which were mainly derived from ovalbumin. Among these peptides, two novel peptides with potent ACE-inhibitory activity were found, with amino acid sequences Arg-Ala-Asp-His-Pro-Phe-Leu and Tyr-Ala-Glu-Glu-Arg-Tyr-Pro-Ile-Leu. Likewise, Davalos *et al.* (2004) reported that enzymatic hydrolysis of crude egg white proteins also resulted in the production of peptides with strong antioxidant activities, and suggested that their combined antioxidant and ACE-inhibitory properties would make a useful multifunctional preparation for the control of cardiovascular diseases, in particular, hypertension.

Ovomucin was also found to inhibit cholesterol uptake *in vitro* by Caco-2 cells, and reduce serum cholesterol in rats, displaying hypocholesterolemic action (Nagaoka *et al.*, 2002).

Egg yolk

The major constituents of egg yolk are proteins and lipids, present mainly in the form of lipoproteins, and can be separated into a granule fraction and plasma fraction (Sugino *et al.*, 1997). The granules contain α - and β -lipovitellins (high-density lipoproteins), phosvitin, and low-density lipoproteins (Burley and Cook, 1961). The plasma can be divided into the low-density lipoprotein fraction and the water-soluble fraction, which contains livetins, which are lipid-free globular proteins, among them γ -livetins, also referred to as immunoglobulin Y (Li-Chan *et al.*, 1995). Egg yolk also contains minerals, carbohydrates, most of which are oligosaccharides bound to protein, as well as pigment (Sugino *et al.*, 1997).

ANTIMICROBIAL ACTIVITY

Although the egg white is the main line of defence against invading microorganisms, a number of egg yolk components have also demonstrated antimicrobial activity. One of the most extensively studied is immunoglobulin (Ig) Y, which has been reviewed in detail elsewhere (Kovacs-Nolan and Mine, 2004a, b). IgY has been produced against a number of bacteria and viruses, and has been shown to bind to and inhibit the infection and disease symptoms, *in vitro* and *in vivo*, of gastrointestinal pathogens (Kovacs-Nolan and Mine, 2004a, b). IgY against *Streptococcus mutans* have been shown to prevent adhesion of the bacteria *in vitro* and *in vivo*, and reduced dental caries development in an animal model (Hatta *et al.*, 1997; Smith *et al.*, 2001; Kruger *et al.*, 2004). In human studies, orally administered anti-*P. aeruginosa* IgY was found to prevent *P. aeruginosa* colonization in the lungs of cystic fibrosis patients, indicating its use as an alternative to antibiotic treatment (Carlander *et al.*, 2000; Kollberg *et al.*, 2003). And the suppression of *H. pylori* infection in humans was achieved/observed following the consumption of drinking yogurt fortified with IgY against *H. pylori* urease enzyme (Horie *et al.*, 2004). Brady *et al.* (2002) found that egg yolk lipoproteins were also important for lipid-mediated antimicrobial activity. The authors found that lipids and lipoproteins, as well as their component fatty acids, oleic and linoleic acid, extracted from egg yolk inhibited the growth of *S. mutans in vitro* (Brady *et al.*, 2003). Finally, egg yolk phosvitin, an iron-binding phosphoprotein, has also demonstrated antibacterial activity against *E. coli* under thermal stress, causing disruption of cells and DNA leakage, suggested to be a result of the synergistic effect of its high metal-chelating ability and high surface activity (Choi *et al.*, 2004).

ANTI ADHESIVE PROPERTIES

Several studies have demonstrated that oligosaccharides in foods such as milk and eggs have an inhibitory effect on the binding of bacteria and viruses to host cells (Sugita-Konishi *et al.*, 2004). The use of natural antimicrobials is a particularly attractive approach, due to the increasing prevalence of microorganisms resistant to, or untreatable by, traditional antibiotic therapy (Kobayashi *et al.*, 2004). Koketsu *et al.* (1995) examined the effect anti-adhesive effect of egg yolk sialyloligosaccharides on viral infection. They found that the oligosaccharide-enriched fraction of egg yolk and sialyloligosaccharide fraction inhibited rotavirus infection *in vitro* in a dose-dependent manner, suggesting that sialic acid plays an important role in binding and inhibition of the virus. They also found that egg yolk sialyloligosaccharides significantly decreased the incidence of rotavirus diarrhoea *in vivo*, in mice (Koketsu *et al.*, 1995) and in rats (Koketsu *et al.*, 1996). More recently, the effects of egg yolk and its components on bacterial adhesion have also been studied. Sugita-Konishi *et al.* (2002) examined the effects of egg yolk sialyloligosaccharides and their derivatives, asialo-yolk-derived sialyloligosaccharides, and sialylglycopeptide, on the binding of *Salmonella enteritidis* and *Escherichia coli*. These compounds inhibited the binding of both organisms to human intestinal cells *in vitro*, and prevented *Salmonella* infection when orally administered to mice. Non-immunized egg yolk powder was also found to eliminate and prevent the intestinal colonization of *S. enteritidis in vitro*, in laying hens when administered at a concentration of 5% in normal feed (Kassaify and Mine, 2004a), and eliminated or significantly reduced colonization of *S. typhimurium*, *Campylobacter jejuni*, and *E. coli* O157:H7, at concentrations of 7.5 and 10% (Kassaify and Mine, 2004b). Furthermore, these pathogens were absent or significantly reduced in internal organs, indicating that the egg yolk

supplement inhibited colonization and invasion, suggested that the observed effects were due to the presence a yet unidentified/novel anti adhesive factor of egg yolk (Kassaify and Mine, 2004a).

ANTIOXIDANT PROPERTIES

Phosvitin has also been recognized as an egg yolk antioxidant protein, acting as an antioxidant by chelating iron ions (Lu and Baker, 1986). Recently, phosvitin and its enzymatic digests were found to protect against iron-catalyzed hydroxyl radical formation, and protect DNA against oxidative damage induced by Fe(II) and peroxide, suggesting that phosvitin may be useful for the prevention of iron-mediated oxidative stress related diseases, such as colorectal cancer (Ishikawa *et al.*, 2004).

NUTRIENT BIOAVAILABILITY

In general, phosvitin has been considered to have no real nutritional value, due to low iron bioavailability and resistance to proteolytic enzymes. However, phosvitin phosphopeptides, derived by tryptic hydrolysis following partial alkaline dephosphorylation have been found to enhance calcium-binding capacity and inhibit the formation of insoluble calcium phosphates (Jiang and Mine, 2000, 2001). Furthermore, phosvitin peptides demonstrated better calcium solubilizing activity than commercial casein phosphopeptides (Jiang and Mine, 2000; Choi *et al.*, 2004).

References

- ABDALLAH, F.B. and CHAHINE, J.M.** (1999) Transferrins, the mechanism of iron release by ovotransferrin. *Eur. J. Biochem.* **263**, 912-920.
- AGUILERA, O., QUIROS, L.M. and FIERRO, J.F.** (2003) Transferrins selectively cause ion efflux through bacterial and artificial membranes. *FEBS Lett.* **548**, 5-10.
- ASAKURA, K., KOJIMA, T., SHIRASAKI, H. and KATAURA, A.** (1990). Evaluation of the effects of antigen specific immunotherapy on chronic sinusitis in children with allergy. *Auris Nasus Larynx* **17**, 33-38.
- BANKS, J.G., BOARD, R.G. and SPARKS, N.H.C.** (1986) Natural antimicrobial systems and their potential in food preservation of the future. *Biotechnol. Appl. Biochem.* **8**, 103-147.
- BARON, F., FAUVEL, S. and GAUTOER, M.** (2000) Behaviour of Salmonella enteritidis in industrial egg white: egg naturally contains factors inhibitory to salmonella growth. In: *Egg Nutrition and Biotechnology*; Sim, J. S.; Nakai, N.; Guenter, W., Eds; CAB International: Oxon, UK, pp. 417-430.
- BRADY, D., GAINES, S., FENELON, L., MCPARTLIN J. and O'FARRELLY, C.A.** (2002) A lipoprotein-derived antimicrobial factor from hen-egg yolk is active against *Streptococcus* species. *J. Food Sci.* **67**, 3096-3103.
- BURLEY, R.W. and VADEHRA, D.V.** (1989) *The Avian Egg, Chemistry and Biology*. John Wiley & Sons, Inc.: New York, New York.
- CARLANDER, D., KOLLBERG, H., WEJAKER, P.E. and LARSSON, A.** (2000) Peroral immunotherapy with yolk antibodies for the prevention and treatment of enteric infections. *Immunol. Res.* **21**, 1-6.
- CHOI, I., JUNG, C., SEOG, H. and CHOI, H.** (2004) Purification of phosvitin from egg yolk and determination of its physiochemical properties. *Food Sci. Biotechnol.* **13**, 434-437.
- COTTERILL, O.J. and GEIGER, G.S.** (1977) Egg product yield trends from shell eggs. *Poultry Sci.*, **56**, 1027-1031.
- DAS, S., BANERJEE, S. and GUPTA, J.D.** (1992) Experimental evaluation of preventative and therapeutic potential of lysozyme. *Chemotherapy* **38**, 350-357.
- DAVALOS, A., MIGUEL, M., BARTOLOME, B., and LOPEZ-FINDINO, R. Davalos, A.** (2004) Antioxidant activity of peptides derived from egg white proteins by enzymatic hydrolysis. *J. Food Prot.* **67**, 1939-1944.
- FAN, X., SUBRAMANIAM, R., WEISSM, M.F. and MONNIER, V.M.** (2003) Methylglyoxal-bovine serum albumin stimulates tumor necrosis factor alpha secretion in RAW 264.7 cells through activation of mitogen-activating protein kinase, nuclear factor kappaB and intracellular reactive oxygen species formation. *Arch. Biochem. Biophys.* **409**, 274-286.
- FUJITA, H., SASAKI, R. and YOSHIKAWA, M.** (1995) Potentiation of the antihypertensive activity of orally administered ovokinin, a vasorelaxing peptide derived from ovalbumin, by emulsification in egg phosphatidylcholine. *Biosci. Biotechnol. Biochem.* **59**, 2344-2345.

- FUJITA, H., USUI, H., KURAHASHI, K. and YOSHIKAWA, M. (1995) Isolation and characterization of ovokinin, a bradykinin B1 agonist peptide derived from ovalbumin. *Peptides* **16**, 785-790.
- GIANSANTI, F., ROSSI, P. MASSUCCI, M.T., BOTTI, D., ANTONINI, G., VALENTI, P. and SEGANTI, L. (2002) Antiviral activity of ovotransferrin discloses an evolutionary strategy for the defensive activities of lactoferrin. *Biochem. Cell Biol.* **80**, 125-130.
- GOLDBERG, J., SHRIKANT, P. and MESCHER, M.F. (2003) In vivo augmentation of tumor-specific CTL responses by class I/peptide antigen complexes on microspheres (large multivalent immunogen). *J. Immunol.* **170**, 228-235.
- HATTA, H., TSUDA, K., OZEKI, M., KIM, M. YAMAMOTO, T., OTAKE, S., HOROSAWA, M. KATZ, J., CHILDERS, N.K. and MICHALEK, S.M. (1997) Passive immunization against dental plaque formation in humans: Effect of a mouth rinse containing egg yolk antibodies (IgY) specific to *Streptococcus mutans*. *Caries Res.* **31**, 268-274.
- HE, X., TSANG, T.C., LUO, P., ZHANG, T., and HARRIS, D.T. (2003) Enhanced tumor immunogenicity through coupling cytokine expression with antigen presentation. *Cancer Gene Ther.* **10**, 669-677.
- HIROTA, Y., YANG, M.P., ARAKI, S., YOSHIHARA, K., FURUSAWA, S., YASUDA, M., MOHAMED, A., MATSUMOTO, Y. and ONODERA, T. (1995) Enhancing effects of chicken egg white derivatives on the phagocytic response in the dog. *J. Vet. Med. Sci.* **57**, 825-829.
- HOLEN, E., BOLANN, B. and ELSAYED, S. (2001) Novel B and T cell epitopes of chicken ovomucoid (Gal d 1) induce T cell secretion of IL-6, IL-13, and IFN-gamma. *Clin. Exp. Allergy* **31**, 952-964.
- HORIE, K., HORIE, N., ABDOU, A.M., YANG, J-O, YUN, S.S., CHUN, C.K., KIM, M. and HATTA, H. (2004) Suppressive effect of functional drinking yogurt containing specific egg yolk immunoglobulin on *Helicobacter pylori* in humans. *J. Dairy Sci.* **87**, 4703-4079.
- IBRAHIM, H.R., SUGIMOTO, Y. and AOKI, T. (2000) Ovotransferrin antimicrobial peptide (OTAP-92) kills bacteria through a membrane damage mechanism. *Biochim. Biophys. Acta* **1523**, 196-205.
- IBRAHIM, H.R., THOMAS, U. and PELLEGRINI, A. (2001). A helix-loop peptide at the upper lip of the active site cleft of lysozyme confers potent antimicrobial activity with membrane permeabilization action. *J. Biol. Chem.* **276**, 43767-43774.
- ISHIKAWA, S., YANO, Y., ARIHARA, K., and ITOH, M. (2004) Egg yolk phosvitin inhibits hydroxyl radical formation from the Fenton reaction. *Biosci. Biotechnol. Biochem.* **68**, 1324-1331.
- JIANG, B. and MINE, Y. (2000) Preparation of novel functional oligophosphopeptides from hen egg yolk phosvitin. *J. Agric. Food Chem.* **48**, 990-994.
- JIANG, B.; and MINE, Y. (2001) Phosphopeptides derived from hen egg yolk phosvitin: effect of molecular size on the calcium-binding properties. *Biosci. Biotechnol. Biochem.* **65**, 1187-1190.
- KASSAIFY, Z.G. and MINE, Y. (2004). Non-immunized egg yolk powder can suppress the colonization of *Salmonella typhimurium*, *E coli* 0157:H7 and *Campylobacter jejuni* in laying hens. *Poultry Sci.*, **83**, 1497-1506.
- KASSAIFY, Z.G. and MINE, Y. (2004). Effect of food protein supplements on *Salmonella enteritidis* infection and prevention in laying hens. *Poultry Sci.*, **83**, 753-760.
- KATO, T., IMATANI, T., MIURA, T., MINAGUCHI, K., SAITOH, E. and OKUDA, K. (2000) Cytokine-inducing activity of family 2 cystatins. *Biol. Chem.* **381**, 1143-1147.
- KOBAYASHI, K., HATTORI, M., HARA-KUDO, Y., OKUBO, T., YAMAMOTO, S., TAKITA, T. and SUGITA-KONISHI, Y. (2004) Glycopeptide derived from hen egg ovomucin has the ability to bind enterohemorrhagic *Escherichia coli* O157:H7. *J. Agric. Food Chem.* **52**, 5740-5746.
- KOKETSU, M., NITODA, T., JUNEJA, L.R., KIM, M., KASHIMURA, N. and YAMAMOTO, T. (1995) Sialyloligosaccharides from egg yolk as an inhibitor of rotaviral infection. *J. Agric. Food Chem.* **43**, 858-861.
- KOKETSU, M., ENOKI, Y., JUNEJA, L.R., KIM, M., and YAMAMOTO, T. (1996) Isolation of sialyloligosaccharides from egg yolk using enzymes and some biofunctional activities of the oligosaccharides isolated. *J. Appl. Glycosci.* **43**, 283-287.
- KOLLBERG, H., CARLANDER, D., OLESEN, H., WEJAKER, P.E., JOHANNESSON, M. and LARSSON, A. (2003) Oral administration of specific yolk antibodies (IgY) may prevent *Pseudomonas aeruginosa* infections in patients with cystic fibrosis: A phase I feasibility study. *Pediatr. Pulmonol.* **35**, 433-440.
- KORPELA, J., SALONEN, E.M., KUUSELA, P., SARVAS, M. and VAHERI, A. (1984) Binding of avidin to bacteria and to the outer membrane porin of *Escherichia coli*. *FEMS Microbiol. Lett.* **22**, 3-10.

- KRUGER, C., PEARSON, S.K., KODAMA, Y., VACCA SMITH, A., BOWEN, W.H., and HAMMARSTRON, L.** (2004) The effects of egg-derived antibodies to glucotransferases on dental caries in rats. *Caries Res.* **38**, 9–14.
- LEE-HUANG, S., HUANG, P.L., SUN, Y., HUANG, P.L., KUMG, H.F., BITHE, D.L. and CHEN, H.C.** (1999) Lysozyme and RNases as anti-HIV components in beta-core preparations of human chorionic gonadotropin. *Proc. Natl. Acad. Sci. U. S. A.* **96**, 2678-2681.
- LI-CHAN, E. and NAKAI, S.** (1989) Biochemical basis for the properties of egg white. *Crit. Rev. Poult. Biol.* **2**, 21-58.
- LI-CHAN, E., POWRIE, W.D. and NAKAI, S.** (1995) The Chemistry of eggs and egg products. In: *Egg Science and Technology*, 4th Edition; Stadelman, W. J.; Cotterill, O. J., Eds.; The Haworth Press, Inc.: New York, New York, pp. 105-175.
- LOSSO, J.N., NAKAI, S. and CHARTER, E.A.** (2000) Lysozyme. In: *Natural Food Antimicrobial Systems*; Naidu, A. S., Ed.; CRC Press, Inc.: New York, New York, pp. 185-210.
- LU, C.L., and BAKER, R.** (1986) Characteristics of egg yolk phosvitin as an antioxidant for inhibiting metal-catalyzed phospholipid oxidations. *Poult. Sci.*, **65**, 2065-2070.
- MATOKA, N., USUI, H., FUJITA, H. and YOSHIKAWA, M.** (1999) A novel anti-hypertensive peptide derived from ovalbumin induces nitric oxide-mediated vasorelaxation in an isolated SHR mesenteric artery. *FEBS Lett.* **452**, 181-184.
- MATOKA, N., YAMADA, Y., USUI, H., NAKAGIRI, R. and YOSHIKAWA, M.** (2001) Designing potent derivatives of ovokinin(2-7), an anti-hypertensive peptide derived from ovalbumin. *Biosci. Biotechnol. Biochem.* **65**, 736-739.
- MIGUEL, M., RECIO, I., GOMEZ-RUIZ, J.A., RAMOS, M. and LOPEZ-FANDINO, R.** (2004) Angiotensin I-converting enzyme inhibitory activity of peptides derived from egg white proteins by enzymatic hydrolysis. *J. Food Prot.* **67**, 1914–1920.
- MINE, Y., MA, F. and LAURIAU, S.** (2004) Antimicrobial peptides released by enzymatic hydrolysis of hen egg white lysozyme. *J. Agric. Food Chem.*, **52**, 1088-1094.
- NAGAOKA, S., MASAOKA, M., ZHANG, Q., HASEGAWA, M. and WATANABE, K.** (2002) Egg ovomucin attenuates hypercholesterolemia in rats and inhibits cholesterol absorption in Caco-2 cells. *Lipids* **37**, 267-272.
- OGURO, T., OHAKI, Y., ASANO, G., EBINA, T. and WATANABE, K.** (2001) Ultrastructural and immunohistochemical characterization on the effect of ovomucin in tumor angiogenesis. *Jpn. J. Clin. Electron Microsc.* **33**, 89-99.
- OTANI, H., and ODASHIMA, M.** (1997) Inhibition of proliferative responses of mouse spleen lymphocytes by lacto- and ovotransferrins. *Food Agric. Immunol.* **9**, 193-202.
- PACOR, S., GAGLIARD, R., DI DANIEL, E., VADORI, M. and SAVA, G.** (1999) In vitro down regulation of ICAM-1 and E-cadherin and in vitro reduction of lung metastases of TS/A adenocarcinoma by a lysozyme derivative. *Int. J. Mol. Med.* **4**, 369-375.
- PELLERGRINI, A., THOMAS, U., BRAMAZ, N., KLAUSER, S., HUNZIKER, P., and VON FELLEBERG, R.** (1997) Identification and isolation of a bactericidal domain in chicken egg white lysozyme. *J. Appl. Microbiol.* **82**, 372-378.
- PELLERGRINI, A., THOMAS, U., WILD, P., SCHRANER, E. and VON FELLEBERG, R.** (2000) Effect of lysozyme or modified lysozyme fragments on DNA and RNA synthesis and membrane permeability of *Escherichia coli*. *Microbiol. Res.* **155**, 69-77.
- PELLERGRINI, A., HULSMIEIER, A., HUNZIKER, P. and THOMAS, U.** (2004) Proteolytic fragments of ovalbumin display antimicrobial activity. *Biochim. Biophys. Acta* **1672**, 76–85.
- SHARON, N., AND OFEK, I.** (2002) Fighting infectious diseases with inhibitors of microbial adhesion to host tissues. *Crit. Rev. Food Sci. Nutr.* **42**, 267–272.
- SAVA, G., CESCIA, V., PACOR, S., and ZABUCCHI, G.** (1991) Observations on the antimetastatic action of lysozyme in mice bearing Lewis lung carcinoma. *Anticancer Res.* **11**, 1109-1113.
- SAVA, G., PACOR, S., DASIC, G., and BERGAMO, A.** (1995) Lysozyme stimulates lymphocyte response to ConA and IL-2 and potentiates 5-fluorouracil action on advanced carcinomas. *Anticancer Res.* **15**, 1883-1888.
- SAVA, G.,** (1996) Pharmacological aspects and therapeutic applications of lysozymes. *EXS* **75**, 433-449.
- SMITH, D.J., KING, W.F. and GODISKA, R.** (2001) Passive transfer of immunoglobulin Y to *Streptococcus mutans* glucan binding protein B can confer protection against experimental dental caries. *Infect. Immun.* **69**, 3135-3142.
- SUGAHARA, T., MURAKAMI, F., YAMADA, Y., and SASAKI, T.** (2000). The mode of actions of lysozyme as an immunoglobulin production stimulating factor. *Biochim. Biophys. Acta* **1475**, 27-34.

- SUGINO, H., NITODA, T. and JUNEJA, L.R.** (1997) General chemical composition of hen eggs. In: *Hen Eggs, Their Basic and Applied Science*; Yamamoto, T.; Juneja, L. R.; Hatta, H.; Kim, M., Eds.; CRC Press, Inc.: New York, New York, pp. 13-24.
- SUGITA-KONISHI, Y., KOBAYASHI, K., SAKANAKA, S., JUNEJA, L.R. and AMANO, F.** (2004) Preventive effect of sialylglycopeptide–nondigestive polysaccharide conjugates on *Salmonella* infection. *J. Agric. Food Chem.* **52**, 5443–5448.
- TANIZAKI, H., TANAKA, H., IWATA, H. and KATO, A.** (1997) Activation of macrophages by sulfated glycopeptides in ovomucin, yolk membrane, and chalazae in chicken eggs. *Biosci. Biotechnol. Biochem.*, **61**, 1883-1889.
- TSUGE, Y., SHIMOYAMADA, M. and WATANABE, K.** (1996) Binding of egg white proteins to viruses. *Biosci. Biotechnol. Biochem.* **60**, 1503-1504.
- TSUGE, Y., SHIMOYAMADA, M. and WATANABE, K.** (1996) Differences in hemagglutination inhibition activity against bovine rotavirus and hen newcastle disease virus based on the subunits in hen egg white ovomucin. *Biosci. Biotechnol. Biochem.* **60**, 1505-1506.
- TSUGE, Y., SHIMOYAMADA, M. and WATANABE, K.** (1997) Structural features of newcastle disease virus- and anti-ovomucin antibody-binding glycopeptides from pronase-treated ovomucin. *J. Agric. Food Chem.* **45**, 2393-2398.
- TSUGE, Y., SHIMOYAMADA, M. and WATANABE, K.** (1997) Bindings of ovomucin to newcastle disease virus and anti-ovomucin antibodies and its heat stability based on binding abilities. *J. Agric. Food Chem.* **45**, 4629-4634.
- VALENTI, P., ANTONINI, G., VON HUNOLSTEIN, C., VISCA, P., ORSI, N and ANTONINI, E.** (1983) Studies of the antimicrobial activity of ovotransferrin. *Int. J. Tissue React.* **5**, 97-105.
- VERDOT, L., LALMANACH, G., VERCROYSE, V., HARTMANN, S., LUCIUS, R., HOEBEKE, J., GAUTHIER, F., and VRAY, B.** (1996) Cystatins up-regulate nitric oxide release from interferon-gamma-activated mouse peritoneal macrophages. *J. Biol. Chem.* **271**, 28077-28081.
- VERDOT, L., LALMANACH, G., VERCROYSE, V., HOEBEKE, J., GAUTHIER, F., and VRAY, B.** (1999) Chicken cystatin stimulates nitric oxide release from interferon-gamma-activated mouse peritoneal macrophages via cytokine synthesis. *Eur. J. Biochem.* **266**, 1111-1117.
- VIDOVIC, D., GRADDIS, T., CHEN, F., SLAGLE, P., DIEGEL, M., STEPAN, L., and LAUS, R.** (2002). Antitumor vaccination with HER-2-derived recombinant antigens. *Int. J. Cancer*, **102**, 660-664.
- WAHN, V.** (2003) Primary immunodeficiencies caused by defects of cytokines and cytokine receptors. In *Methods in Molecular Biology: Cytokines and Colony Stimulating Factors: Methods and Protocols*; Korholz, D.; Kiess, W., Eds.; Humana Press Inc.: Totowa, New Jersey, Vol. 215, pp. 3–12.
- WATANABE, K., YSUGE, Y., SHIMOYAMADA, M., OGAMA, N. and EBINA, T.** (1998) Antitumor effects of pronase-treated fragments, glycopeptides, from ovomucin in hen egg white in a double grafted tumor system. *J. Agric. Food Chem.* **46**, 3033-3038.
- XIE, H., HUFF, G.R., HUFF, W.E., BALOG, J.M., and RATH, N.C.** (2002) Effects of ovotransferrin on chicken macrophages and heterophil-granulocytes. *Dev. Comp. Immunol* **26**, 805-815.
- YAMADA, Y., MATOBA, N., USUI, H., ONISHI, K., and YOSHIKAWA, M.** (2002) Design of a highly potent anti-hypertensive peptide based on ovokinin(2-7). *Biosci. Biotechnol. Biochem.* **66**, 1213-1217.
- YOSHIKAWA, M., and FUJITA, H.** (1994) Studies on the optimum conditions to utilize biologically active peptides derived from food proteins. In: *Developments in Food Engineering*; Yano, T; Matsuno, R.; Nakamura, K., Eds.; Blackie Academic and Professional: New York, New York, pp. 1053-1055.