Evaluation of Larval Development of *Dirofilaria immitis* in Different Populations of *Aedes aegypti* and *Aedes albopictus*

Gílcia Aparecida de Carvalho¹*, Rafael Trindade Maia², Rafael Antonio Nascimento Ramos¹, Carlos Fernando Salgueiro da Andrade³, Maria Aparecida da Gloria Faustino³, Leucio Câmara Alves³

¹Unidade Acadêmica de Garanhuns, Universidade Federal Rural de Pernambuco, Garanhuns, Brazil
²Centro de Desenvolvimento Sustentável do Semiárido-Unidade Acadêmica de Educação do Campo, Universidade Federal de Campina Grande, Sumé, Brazil
³Departamento de Medicina Veterinária, Universidade Federal Rural de Pernambuco, Recife, Brazil
⁴Dipartimento di Medicina Veterinaria, Università degli Studi di Bari, Valenzano, Italy
⁵Universidade Estadual de Campinas, Campinas, Brazil

Email: *gilciasilva@yahoo.com.br*

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ABSTRACT

*Dirofilaria immitis* is an important nematode parasite commonly known as heartworm. This filarioid is transmitted by culicid vectors and primarily affects dogs, but other animals may also become infected, such as wild carnivores, cats and humans. The aim of the present study was to assess the development of *D. immitis* larvae in different culicid populations under laboratory conditions. Adult females of populations of *Aedes aegypti* from the city of Recife (P1), the city of Campinas (P2) and the Rockefeller strain from the Centers for Disease Control (P3) and one population of *Aedes albopictus* from Recife (P4) were fed for two hours with infected dog blood containing 2,000 microfilariae/ml of *D. immitis*. After artificial feeding, the specimens were maintained under controlled conditions. Ten females from each population were dissected daily over 14 days. The infection ratio and vector efficiency index were calculated and *D. immitis* development from L1 to L3 was assessed. The larvae in P1, P2, P3 and P4 reached the third stage in 11, 10, 14 and 9 days, respectively. The vector efficiency index was 53.8%, 20.0%, 7.4% and 25.2% in P1, P2, P3 and P4, respectively. The findings demonstrate that *D. immitis* larvae develop in all culicid populations studied herein. Based on mosquito mortality, development time and VEI the *A. albopictus* population from Recife (P4) demonstrated the best performance as vector. This is the second report of *D. immitis* development in *A. albopictus* from Brazil. The present data reinforce the role of this species as vector of *D. immitis* in an area where greater importance has long been given to *A. aegypti*.

Keywords: Culicid; Vector; Heartworm; Dog; Experimental Infection

1. Introduction

Dirofilariasis is an important parasitic disease caused by the nematode *Dirofilaria immitis*, also known as heartworm [1]. This filarioid primarily affects dogs, but infection has also been reported in cats [2,3], wild carnivores [4,5] and humans [6]. In this later host, infection is characterized by the presence of pulmonary nodules (the so-called “coin lesion”) and no apparent symptoms [7].

Transmission occurs through mosquito vectors, which are the most important vectors that belong to the genera *Culex, Aedes* and *Anopheles* [8,9]. Briefly, first stage larvae (L₁) are taken up by blood-sucking female mosquitoes and develop through to the infective final larvae stage (L₃) in approximately 14 days [10]. The vector competence of a culicid population is the primary factor for *D. immitis* transmission in an endemic area. However, characteristics such as the immune reaction, environmental conditions (e.g., temperature and humidity) and genetic traits among *D. immitis* populations may be factors which affect the vector capacity of culicids [11].
In Brazil, studies on *D. immitis* vectors are scarce. In the southeastern region of the country, *Aedes scapularis* and *Aedes taeniorhynchus* have been shown to be the main vectors, while *Culex quinquefasciatus* seems to have secondary importance [2]. Also in this region (state of Rio de Janeiro), an experimental study demonstrated *Aedes aegypti* as a potential vector of *D. immitis* [12]. In the northeastern region *Cx. quinquefasciatus* and *A. taeniorhynchus* have been found naturally infected [13], but little is known regarding the epidemiological importance of these species, especially the latter. Unfortunately, the vector competence of two species well established throughout Brazil (*A. aegypti* and *A. albopictus*), has been poorly studied.

Therefore, the aim of the present study was to assess the development of *D. immitis* larvae in three populations of *A. aegypti* and one of *A. albopictus* from different geographic areas under laboratory conditions.

2. Material and Methods

2.1. *D. immitis* Microfilariae

Blood containing microfilariae was obtained from a dog positive for *D. immitis* in the microscopic analysis (see below). The dog was a three-year-old male that lived in the metropolitan region of Recife (7°45′0″S and 34°51′0″W), state of the Pernambuco, Brazil.

2.2. Microscopic Diagnosis (Modified Knott Test)

The microscopic analysis was performed using the modified Knott test [14]. Briefly, 1 ml EDTA blood was mixed with 9 ml of distilled water and centrifuged for 5 min at 300 × g. The supernatant was discarded, leaving 1 ml of solution to which two drops of methylene blue solution were added. The sediment was transferred to glass slides, covered with coverslip and examined under a light microscope at different magnifications. All filarioid larval found were morphologically identified as *D. immitis*.

2.3. Mosquito Populations and Experimental Infection

Four different mosquito populations were used: *A. aegypti* from the city of Recife, Brazil (P1), *A. aegypti* from the city of Campinas, Brazil (P2), *A. aegypti* Rockefeller strain from the Centers for Disease Control in the USA (P3) and *A. albopictus* from Recife (P4).

A total of 4,800 female mosquitoes (3,600 for the test populations and 1,200 for the control) aged from three to seven days were used [15]. The artificial blood meal was performed as previously described [10]. The test mosquitoes were fed for two hours with the infected blood, which was previously assessed as containing 2,000 microfilariae/ml. Non-infected blood was offered to the control group. After feeding, the mosquitoes were maintained under controlled temperature (28°C ± 2°C) and relative humidity (>70%).

2.4. Mosquito Dissection and Microscopic Examination

For the detection of *D. immitis* larvae and developmental stages, ten mosquitoes from each population were dissected daily. On Day 14, all remaining living mosquitoes were also dissected. The mosquitoes were fixed on slides containing a drop of 0.9% physiological saline solution, dissected with a sterile scalpel and immediately examined under a light microscope (Olympus BX41 TF) at different magnifications. All stages of *D. immitis* larvae were morphologically identified [16].

2.5. Data Analysis

The infection ratio (IR) [17] and vector efficiency index (VEI) were calculated [18]. Linear regression was used to determine the influence of the ingestion of *D. immitis* microfilariae on mosquito mortality. Analysis of variance was used to compare larval development up to the infectious stage in the female mosquitoes as well as mortality in the different populations. The BioStat program (version 2.0) was employed for the statistical analysis [19].

3. Results

After feeding, the mosquitoes exhibited a similar rate of engorgement: 97.6%, 91.5%, 97.6% and 93.3% in P1, P2, P3 and P4, respectively, and 96.6% in the control group. *Table 1* displays the mean ingestion of microfilariae in each population. Microfilaria ingestion was statistically similar among P1, P3 and P4 (*P* > 0.05), whereas P2 ingested a significantly smaller number of microfilariae (*P* < 0.01).

Throughout the study period, mosquito mortality was greater in P1 than P2, P3 and P4 (*P* < 0.01). However, this higher mortality rate was not influenced by the number of microfilariae ingested (*F* = 0.6899; *P* > 0.05). Two peaks in mosquito mortality were found in all populations, one on the second day post-infection and one after the detection of L₃. At the end of the study, overall mortality in the control group was 3.4%.

The infection rate among the populations ranged from 11.6% to 17.5% (*Table 2*). This parameter was not influenced by the proportion of engorged females (*F* = 12.8754; *P* > 0.05), but was affected by the number of microfilariae ingested (*F* = 141.3808; *P* < 0.01). *D. immitis* larval development from L₁ to L₃ occurred in all infected populations. In P1, L₁ was found on Day 3 post-infection, L₂ on Day 7 and a low number of L₃ (*n* = 7) were retrieved on Day 11. In P2 and P3, L₁ on Day 6 post-infection, L₂ was found on Day 8 and L₃ was found
on Days 10 and 14, respectively. Melanized larvae and few larvae remaining as \( L_1 \) were detected at the end of the study in all populations. \( D. \) immitis larvae exhibited a comparatively short development time when infecting \( A. \) albopictus (P4), with larvae reaching \( L_1, L_2 \) and \( L_3 \) at Days 2, 5 and 9, respectively.

The number of \( L_3 \) obtained 14 days post-infection (Table 1) was not influenced by the number of microfilariae ingested \((F = 0.0415; P > 0.05)\). Both mortality peaks occurred during the first days. Interestingly, this parameter was not influenced by the number of microfilariae ingested \((F = 0.6899; P > 0.05)\). Both mortality peaks occurred during critical post-infection times: first during the penetration of \( L_1 \) into the Malpighian tubule cells (between Days 1 and 2 post-infection) and soon after the detection of \( L_3 \). In both events, Malpighian tubule cells were destroyed, with the magnitude of the destruction depending on the quantity of larvae, causing death in P1. As previous states \([21]\) an efficient mosquito vector should survive independently of the number of ingested microfilariae thus, P1 may exhibit a comparatively low degree of vector competence.

\( D. \) immitis development was observed in all populations studied herein, with the maximum VEI \((53.8\%)\) found for \( A. \) aegypti from Recife (P1). However, this population may not be considered an efficient vector due to the high mortality rate observed \((70.7\%)\). The minimal differences in development time found among the populations of \( A. \) aegypti were expected, since the same nematode strain was used for all populations. Previous studies have been reported different development times for this filarioid when different strains of nematode and mosquitoes are used \([16,22]\).

Among the four populations studied, one merits particular attention, \( A. \) albopictus from Recife (P4) exhibited low mortality and a considerable VEI \((25.2\%)\). Furthermore, larvae reached the third infective stage in only nine days. \( A. \) albopictus is reported to be the primary potential vector of \( D. \) immitis in Italy \([23]\). The importance of this mosquito as a vector for \( D. \) immitis in Brazil has been assessed in a single study \([12]\). Nonetheless, its vector capacity has interesting epidemiological implications. \( A. \) albopictus is considered a secondary vector of human arboviruses due to its preferential feeding on animals rather than humans, unlike the highly anthropophilic behavior of \( A. \) aegypti \([24]\). Based on feeding preference \( A. \) albopictus may have a major veterinary importance for \( D. \) immitis transmission among animals, which should be studied better. It is important stress that the \( A. \) albopictus population evaluated herein proved to be susceptible to infection and allowed \( D. \) immitis larvae development. However, further studies evaluating the vector capacity of other populations of this species in different regions of Brazil should be carried out.
5. Conclusion

In conclusion, *D. immitis* larvae developed in all mosquito populations studied. Based on factors as mortality, development time and VEI, the *A. albopictus* population from Recife demonstrated the best performance as vector. This study is the second description of the development of *D. immitis* in *A. albopictus* in Brazil. Moreover, the findings suggest that this culicid species may perform an important role as a vector for *D. immitis* in an area (i.e., Brazil) where greater importance has long been attributed to *A. aegypti*. Field studies should be carried out to clarify the real importance of this culicid species in the transmission of *D. immitis* in Brazil.

REFERENCES


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